

Impact of *Aeromonas veronii* Infection in Freshwater Fishes and Its Threat to Mankind: A Review

¹Kushal Thakur, ²Akriti Choudhary, ³Amit Kumar Sharma, ⁴Bhavna Brar, ⁵Danish Mahajan, ⁶Rajinder Jindal, ⁷Devender Singh and ⁸Rakesh Kumar*

Author's Affiliation:

^{1,2,3,4,5,8}Department of Animal Sciences, School of Life Sciences, Central University of Himachal Pradesh, District Kangra, Himachal Pradesh 176 206, India.

⁶Aquatic biology lab, Department of Zoology, Panjab University, Chandigarh 160 014, India

⁷Department of Higher education Himachal Pradesh

*Corresponding author:

Dr. Rakesh Kumar

Department of Animal Sciences, School of Life Sciences, Central University of Himachal Pradesh, District Kangra, Himachal Pradesh 176 206, India

E-mail: drthakurcuhp@gmail.com,
drthakurcuhp@hpcu.ac.in

ABSTRACT:

The emerging pathogen *Aeromonas veronii* is extensively dispersed and can infect fishes, humans and animals. It is a broadly distributed gram-negative, facultatively anaerobic bacterium. It causes Epizootic Ulcerative Syndrome (EUS) and fish sepsis in several freshwater fishes such as tilapia, catfish, carp, and loach etc. The high mortality rates among commercial fish species leads to huge financial losses to the aquaculture sector. Antibiotics were once a successful preventative measure in this regard. However, the widespread use of antibiotics in recent years has resulted in antibiotic resistance. Probiotics, vaccines, and phytochemicals are some of the control measures that can be used in place of antibiotics. The present review will provide information about overview of *A. veronii* infection in fishes, its effect on both human and animal wellbeing, virulence, and its management approaches in aquaculture.

Keywords:

Bacterial infection, *Aeromonas veroni*, Aquaculture fishes, Host-pathogen interaction.

Received on 05.06.2024

Revised on 15.10.2024

Accepted on 25.11.2024

How to cite this article: Thakur K., Choudhary A., Sharma A.K., Brar B., Mahajan D., Jindal R., Singh D. and Kumar R. (2024). Impact of *Aeromonas veronii* Infection in Freshwater Fishes and Its Threat to Mankind: A Review. *Bulletin of Pure and Applied Sciences-Zoology*, 43A (2), 364-382.

1. INTRODUCTION

Aquaculture is one of the swiftly expanding food-producing industries, accounting for over 40% of global fish food production. It is spreading, expanding, and intensifying in nearly every region of this world. Aquaculture is more diversified than other agricultural sectors in terms of species, feeds, production methods, illnesses, products, and organisational structures (Yue & Shen, 2022). With the growing world population, consumer interest in aquaculture is expected to exceed 62% of total global production by 2030 (Ahmad *et al.*, 2021). However, the production of aquaculture is being

hampered by a number of developing diseases caused by bacteria, viruses, fungus, amoebas, oomycetes, and other ectoparasites. Bacteria can survive in any aquatic ecosystem in the absence of their host, so bacterial disease has become an important obstacle to aquaculture (Klesius & Pridgeon, 2011).

Aeromonas which is a member of family Aeromonadaceae, is one such developing bacterial pathogen in aquaculture, inflicting harm to the health of freshwater species through acute or chronic illnesses (Thakur *et al.*, 2022). The *Aeromonas* causes mass mortality in fishes that cause financial losses in aquaculture sector

(Kaur *et al.*, 2020; Yu *et al.*, 2010). *Aeromonas* has a wide dispersion and had been obtained from various sources, including fresh water, vegetables, soil, fruits, processed food and sewage (Barger *et al.*, 2021). This genus of bacteria is recognized to naturally exist in aquatic habitats and commonly known as opportunistic pathogens (Lü *et al.*, 2016; Barger *et al.*, 2021). The *Aeromonas* lateral flagella are crucial for boosting cell attachment, biofilm formation, and invasion which help the bacteria to survive in harsh environments and develop pathogenicity (Gavin *et al.*, 2003). There are 36 species in the *Aeromonas* genus, including *A. veronii*, *A. sobria*, *A. caviae*, and *A. hydrophila* which are regarded as emerging pathogens, not only to fishes, amphibians, and reptiles, but also humans (Wahli *et al.*, 2005; Yu & Park., 2008; Fernández-Bravo & Figueras, 2020).

2. OVERVIEW OF AEROMONAS VERONII IN FRESHWATER FISHES

Oreochromis niloticus, also referred to as the aquatic chicken or the Nile tilapia, is one of the world's fastest-growing aquaculture species. Global tilapia production was projected to be 4.5 million tonnes in 2014, and by 2030, it is expected to reach 7.3 million tonnes, providing the majority of developing nations with an affordable source of protein (Bacharach *et al.*, 2016). In Egyptian fish farms, *Aeromonas veronii* appears to be the primary cause of mass mortality among *Oreochromis niloticus* (Latif *et al.*, 2019). Red tilapia also showed acute mortality due to *A. veronii* (Dong *et al.*, 2015). It was found that TiLV and *A. veronii* spontaneously co-infect a Malaysian red hybrid tilapia (*O. niloticus* × *O. mossambicus*). However, the issue might become worse if TiLV and other bacteria co-infected synergistically (Amal *et al.*, 2018). In juvenile *O. niloticus*, *A. veronii sobria* causes anorexia, abdominal bloating, skin pigmentation, bleeding spots, rotting of the tail and erosion of the caudal fin with a prevalence of 99.80% (Li & Cai, 2011).

Yellow catfish (*Pelteobagrus fulvidraco*) and channel catfish (*Ictalurus punctatus*) are commercially important freshwater-farmed species with a production rate of over 834 thousand tons in 2019 (Yu *et al.*, 2020). These

species of catfish experienced widespread fatalities and epidemics of ulcerative conditions showing symptoms such as ulcerations in the skin, ascites, bleeding, hemorrhagic septicemia due to *A. veronii* (Zhai *et al.*, 2023). *Aeromonas veronii* has been found to be the cause of channel catfish mortality in Vietnam (Hoai *et al.*, 2019). In China and the Indo-Pakistan region, *A. veronii* was isolated from diseased catfish (Cai *et al.*, 2012). The primary signs of disease in farmed channel catfish have been recorded as skin ulcers and bleeding (Gui *et al.*, 2018). When ulcers develop on the skin of the fish due to *A. veronii*, some external bacteria, such as *Kurthia*, (*Kurthia gibsonii*) have also been isolated from infected mucosa in both diseased yellow and channel catfish (Preena *et al.*, 2019; Zhai *et al.*, 2023).

The common carp (*Cyprinus carpio*) is regarded as an important aquaculture species in many European and Asian countries because of its high adaptive capability to both environment and food (Parkos & Wahl, 2014; Choudhary *et al.* 2023; Thakur *et al.*, 2025). In some European countries, common carp accounts for more than 80% of total fish farming (Rahman, 2015). *Aeromonas veronii* has been identified as the causative agent of disease outbreaks and mass mortality in *Cyprinus carpio* in Korea (Yu *et al.*, 2010). Chen *et al.* (2019) also discovered *A. veronii* in diseased crucian carp in a Chinese fish farm (Chen *et al.*, 2019). A variety of clinical signs, such as gill congestion, hyphema, cutaneous hemorrhage, hemorrhages in the fins and operculum, abdominal swelling, abdominal congestion, hepatic portal redness, skin lesions (erythrodermatitis), intestinal swelling, dropsy symptoms are frequently present in *A. veronii* infected crucian carp (Yu *et al.*, 2010; Chen *et al.*, 2019). In addition to crucian carp, grass carp, and cyprinid fish also exhibit these symptoms when infected with *A. veronii* (Ran *et al.*, 2018).

Numerous studies have documented the tragic deaths and huge financial losses of ornamental fish from bacterial infections (Sahoo *et al.*, 2016; Lewbart, 2001). The most popular ornamental fish species worldwide are goldfish (*Carassius auratus*), zebra danio (*Danio rerio*), neon tetra (*Paracheirodon innesi*), angelfish (*Pterophyllum* spp.), discus (*Symphysodon* spp.) and guppy

(*Poecilia reticulata*). Over 14% of the traded fish is made up of guppy and zebra danio. Additionally, koi fish (*Carassius auratus*), which are primarily exported from Japan, make up 10% of world trade. *Aeromonas veronii* is a significant bacterial pathogen and also a symbiont in the digestive tract of zebrafish (Bates *et al.*, 2006; Silver *et al.*, 2011). In ornamental fish farms, the prevalence of skin ulcerative illness caused by *A. veronii*, which is characterised by internal and external bleeding

is increasing (Sun *et al.*, 2016). Strongly virulent to ornamental fish, *A. veronii* causes glomerulus hemorrhage, necrosis of the hepatic cell, leukocyte infiltration of gill filaments, and skin ulcerative syndrome (Song *et al.*, 2017). Goldfish farms in India suffered significant losses due to *A. veronii* infection. These infected goldfish displayed symptoms such as anorexia, a swollen abdomen, tail rot, and cutaneous haemorrhage (Shameena *et al.*, 2020).

Table 1: Different species of freshwater fishes infected by *A. veronii*

S. No.	Affected Species	Common name	Diseased caused	symptoms	Reference
1	<i>Oreochromis niloticus</i>	Nile tilapia	Hemorrhagic septicemia	Anorexia, lethargy, detached scales, abdominal bloating, skin pigmentation, bleeding spots, rotting of the tail and erosion of the caudal fin	Li and Cai, 2011
2	<i>Pelteobagrus fulvidraco</i>	Yellow catfish	Epizootic ulcerative syndrome	Abdominal enlargement and body surface ulcers	Fu <i>et al.</i> , 2021
3	<i>Ictalurus punctatus</i>	Channel catfish	Hemorrhagic septicemia	Skin ulcers and bleeding	Yang <i>et al.</i> , 2017; Gui <i>et al.</i> , 2018
4	<i>Ctenopharyngodon idella</i>	Grass carp	Bacterial septicemia	Abdominal ascites, bleeding from the skin and internal organs	Wu <i>et al.</i> , 2021
5	<i>Cyprinus carpio</i>	Common carp	Motile Aeromonad Septicemia (MAS)	Abdominal congestion, hepatic portal redness, abdominal distention, skin lesions and abdominal swelling	Ran <i>et al.</i> , 2018
6	<i>Carassius auratus gibelio</i>	Crucian carp	Haemorrhagic septicemia	Gill congestion, hyphema, cutaneous hemorrhage,	Chen <i>et al.</i> , 2019

				intestinal swelling, dropsy symptoms	
7	<i>Carrasius auratus</i>	Goldfish	Epizootic ulcerative syndrome	Skin lesions, anorexia, swollen abdomen, exophthalmia, fin and tail rot, and cutaneous haemorrhages	Song <i>et al.</i> , 2017; Shameena <i>et al.</i> , 2020
8	<i>Odontobutis potamophila</i>	River snakehead	Epizootic ulcerative syndrome	Hemorrhage on fins, ulceration on the abdomen	(Liu <i>et al.</i> , 2022)
9	<i>Astronotus ocellatus</i>	Oscar	Motile Aeromonad Septicemia (MAS)	Infectious abdominal dropsy, anorexia, lethargy, scale protrusion and petechial haemorrhage	Sreedharan <i>et al.</i> , 2011
10	<i>Myxocyprinus asiaticus</i>	Chinese Sailfin Sucker	Motile Aeromonad Septicemia (MAS)	Rotting fins, red ascitic fluid, liver and kidney swelling, hyperaemia, a black swollen spleen, pale gills, cloacal pore swelling and intestinal inflammation.	Li <i>et al.</i> , 2019
11	<i>Ophiocephalus argus</i>	Snakehead fish	Enteritis Epizootic ulcerative syndrome(EUS)	Enteritis- Loss of appetite, red blotches on abdomen, congested fins, swollen anus, and ascites. EUS- Ulcers on skin, fins, and abdomen, lethargy, swollen abdomen and bleeding.	Wang <i>et al.</i> , 2020; Zheng <i>et al.</i> , 2012
12	<i>Anabas testudineus</i>	Climbing perch	Epizootic ulcerative syndrome(EUS)	Swollen abdomen, ulcers on skin, skin pigmentation,	Ehsan <i>et al.</i> , 2023

				bleeding spots, rotting of the tail	
13	<i>Micropterus salmoides</i>	Largemouth bass	Epizootic ulcerative syndrome(EUS)	Gill hemorrhage, fin hemorrhage, dorsal and abdomen ulcers , and rotting fins	Pei <i>et al.</i> , 2021
14	<i>Labeo rohita</i>	Rohu	Motile Aeromonad Septicemia (MAS)	Haemorrhage, rotting fins and gross lesions.	Tyagi <i>et al.</i> , 2022
15	<i>Paramisgurnus dabryanus</i>	Loach	Hemorrhagic septicemia	Ulcers on the body surface, hemorrhages under the skin, darkening of liver, necrosis of liver, lesions in the liver, spleen, kidney and muscle	Du <i>et al.</i> , 2020
16	<i>Procambarus clarkii</i>	Crayfish	Motile Aeromonad Septicemia (MAS)	Weakness of limbs, slow movements, lethargy, gill rot, and weak stress ability	Yang <i>et al.</i> , 2022

3. A. VERONII AS A PUBLIC HEALTH ISSUE

Zoonotic disease are those infectious disease which are transmitted from an animal host to a human host (Han *et al.*, 2016). The majority of fish-related zoonotic diseases spread to humans by eating raw or improperly cooked fish and drinking water contaminated with sick fish (Ziarati *et al.*, 2022). This pathogen can still invade through bruises, which can occur while handling fish aquaria, or while dealing with diseased fish (Gauthier, 2015). The fish pathogen *A. veronii* can also infect humans with weak immunity causing gastroenteritis, wound infections, and septicemia (Fernández-Bravo & Figueras, 2020). These patients may also experience some other symptoms like nausea, abdominal pain, diarrhoea, and vomiting (Vila *et al.*, 2003). Potential health risk arises from the abundance of *Aeromonas* in foods and drinking water. In warm weather, humans are most likely to contract stomach illnesses from *A. veronii* (Yuwono *et al.*, 2021). *Aeromonas veronii*

prevalence varies geographically and is associated with poor sanitation practices in less developed areas (Tsai *et al.*, 2015). Mohan *et al.* (2017) evaluated 50 samples of diarrheal stools from 1,595 patients to identify *Aeromonas*; the biochemical analysis of 35 strains revealed that *A. biovar sobria* and *A. veronii biovar veronii* were the most prevalent species (Mohan *et al.*, 2017). *Aeromonas veronii* was found in abundance in 193 of the 4,529 diarrheal patient samples collected from 16 institutions in Shanghai, China (Li *et al.*, 2015). According to a study conducted in Taiwan, 76 cases per million individuals occurred between 2008 and 2010 (Wu *et al.*, 2014). At Galdakao-Usansolo Hospital in Spain from January 2015 to December 2017, 98 patients were enumerated who had positive stool cultures for *A. veronii*, giving an estimated incidence of 32 cases for every 105 individuals annually (Elorza *et al.*, 2020). According to a study of 109 clinical isolates of *Aeromonas* from patients with diarrhoea, *A. veronii* was the most common species in both Mexico and Spain

(Aguilera-Arreola *et al.*, 2007). Both healthy individuals and individuals on immunosuppressive medications have had genitourinary tract infections brought on by *A. veronii* (Chao *et al.*, 2012). Although a majority of patients recover without medical intervention, those who have severe symptoms and persistent illnesses frequently need hospitalization and antibiotic therapy (Chen *et al.*, 2015).

4. ANIMAL HEALTH CONCERNS RELATED TO *A. VERONII*

Aeromonads can frequently be retrieved from vertebrates as well as other hosts like insects. Pigs, poultry, cattle, sheep, buffalo, and fish are just a few of the food animals and products that have been shown to have *A. veronii* (Gowda *et al.*, 2015). A number of aquatic fauna can be infected by *A. veronii* which mainly causes organ bleeding, skin ulcers, and severe ascites. Various *M. nipponensis* farms, experienced fatalities from June to October of the year 2019 in Xinghua county, Jiangsu province, China. These sick *M. nipponensis* had red gills, weakness, and a poor appetite as symptoms of their illness. Results showed that *A. veronii* was the pathogen responsible for *M. nipponensis*'s high mortality rate (Gao *et al.*, 2020). *Macrobrachium rosenbergii* could die in large numbers as a result of the *A. veronii* (Sung *et al.*, 2000). In the Al-Ahsaa area, marsh frogs have experienced outbreaks of Ulcerative Syndrome brought on by *A. veronii* *bv. veronii*, which is especially harmful to *P. ridibundus* larvae (Khalifa *et al.*, 2021). *Aeromonas veronii* caused outbreaks of respiratory disease in a sheep breeding farm in Shaanxi, China, showed 2.53% (15/594) of sheep with respiratory (clinical) symptoms like dyspnea, nasal discharge, wet cough, fever, and progressive emaciation disease (Miao *et al.*, 2023).

5. PATHOGENICITY

Aeromonas veronii is known to cause Epizootic Ulcerative Syndrome (EUS) or Motile Aeromonad Septicemia (MAS) characterized by glomerulus hemorrhage, swollen abdomen, skin pigmentation, fin and tail rot, dropsy symptoms, necrosis of the hepatic cell, leukocyte infiltration of gill filaments, and skin ulceration (Li & Cai,

2011; Song *et al.*, 2017; Fu *et al.*, 2021). Previous research have shown that a number of connected variables help to make *A. veronii* pathogenic. There are recognized virulence characters, including hemolytic and cytotoxic activity, toxins, iron ion acquisition systems, proteases, outer membrane proteins, secretion systems, quorum, and motility-related factors (Qin *et al.*, 2022). Another link between pathogenicity and virulence is the presence of the cytotoxic enterotoxins, aerolysin, glycerophospholipid: cholesterol acyltransferase, serine, lipase, and protease genes which were identified in an infected crucian carp (Hossain *et al.*, 2018; Chen *et al.*, 2019). The most frequent virulence genes identified in an infected tilapia are lipase (lip), elastase (ahp), hemolysin (hly), and aerolysin (aer) (Gohary *et al.*, 2020). These virulent genes contribute to the pathogenicity of *A. veronii* by encoding proteins and toxins (Zhu *et al.*, 2022). The enterotoxin-producing genes act and alt, aerA, as well as hemolytic toxin-producing genes hlyA, all have an important role to play in the virulence of *A. veronii* (Sreedharan *et al.*, 2013). The aer gene is a crucial gene linked to aerolysin while the fla gene is crucial for the ability of motility and cell adhesion (Abrami *et al.*, 2003; Sen and Rodgers, 2004). The gene hly is important for the cytotoxic effects including erythrocyte lysis whereas the vipB gene plays an important role in virulence, adhesion ability, motility, and resistance to oxidative stress (Gao *et al.*, 2013; Song *et al.*, 2023).

6. HOST-PATHOGEN INTERACTION

Various innate immune system tissues and functions have been studied in *M. salmoides*, a fish species with an adaptive immune system that can generate a targeted antibody response against infections (Zhu *et al.*, 2022). Specifically, hemolysin produced by *A. veronii* caused injury to the host spleen. Several immune-related signalling pathways are favoured in the host during infection, including Toll-Like Receptor Signaling Pathway, T Cell Receptor Signaling Pathway, and Cytokine-Cytokine Receptor Interaction (Qin *et al.*, 2023). Figure 1 show the various pathways involved in the host's response towards infection. Toll like receptor pathway with the Tumor necrosis factor (TNF),

Retinoic acid-inducible gene (RIG)-like receptor (RLR) and nuclear factor kappa B (NF- κ B) are involved in inflammatory response generated from the host. To begin innate immune responses, TLRs are very essential and are the first Pattern recognition receptor (PRRs) to be described. It consists of three components: a transmembrane region, a C-terminal cytoplasmic TIR domain that contacts adaptor molecules, and an N-terminal ectodomain with LRR (Leucine-rich repeat) that is necessary for PAMP (Pathogen-associated molecular pattern) recognition (Li *et al.*, 2019). IL15 is a key molecule that regulates the growth of dendritic cells, NK cells, and T-cells as well as taking part in some immune-related signal transmission pathways, which plays a significant role in innate and adaptive immunity (Bae *et al.*, 2013).

In addition to being the main antibody in the primary reaction, IgM plays a crucial role in the fish's adaptive immune response (Defoirdt *et al.*, 2011). TNF- α and IL-1 β can be released during apoptosis, which is induced by the hypoxia-inducible factor (HIF) (Zhang *et al.*, 2021). After infection with *A. veronii*, Zhu *et al.* (2022) discovered that there was a significantly higher level of IgM expression in the liver and spleen as well as higher levels of HIF-1 expression. Caspase-3 is a vital executory enzyme and apoptosis's last effector (Zhu *et al.*, 2022). The host immunological defence system was activated and these immune-related genes were impacted by *A. veronii*, providing a plausible basis for host-pathogen interactions (Zhang *et al.*, 2019).

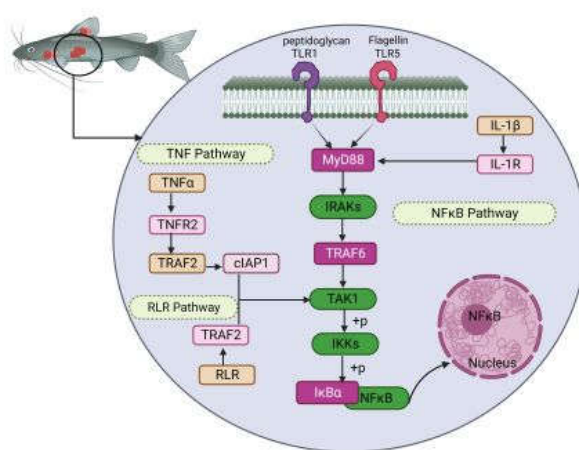


Figure 1: TLR pathway that induces the inflammatory response in the host.

TLR1 and TLR5 are located on the cell surface. TLR1 receives peptidoglycan while TLR5 receives flagellin produced by *A. veronii*. MyD88 is an adaptor molecule that interacts with IRAK-4, which has active IRAK-1 and IRAK-2. The IRAKs then separate from MyD88 and engage with TRAF6, TRAF6 will respond to TAK1 and IKK, after the deduction of immune signals from MyD88 to TRAF6. Activated IKK phosphorylates I κ B α which is degraded via ubiquitin mediation, causing dissociation of NF- κ B, which activates the NF- κ B pathway. The activated NF- κ B binds to target genes, proinflammatory cytokines genes (TNF- α , IL-1 β),

and I κ B α after it translocated to the nucleus. TNF α and IL-1 β production are both strengthened by NF- κ B activation, and both of them in turn activate NF- κ B via TNFRs and IL-1R. (Modified Li *et al.*, 2019)

7. CONTROL AND MANAGEMENT

Fish grown in ponds with recirculation are commonly affected by infections caused by *A. veronii* bacteria, are managed using antibiotics (Saavedra *et al.*, 2004). As a result, antibiotic tolerance and antimicrobial residues could

consequently emerge in aquaculture (Vivekanandhan *et al.*, 2002). Apart from being chemical pollutants, antibiotics also impose a selection pressure that promotes the spread of various antibiotic resistance genes (ARG), endangering human health (Kemper, 2008). Tekedar *et al.* (2020) reported resistance genes in *A. veronii* including chloramphenicol and florfenicol resistance gene (*floR*), beta-lactamase resistance genes (*imiS* and *ampS*), macrolide resistance genes (*mcr-3* and *mcr-7.1*), acetyltransferase genes, tetracycline resistance genes [*tet(34)*, *tet(35)*, and *tet(E)*], and streptogramin B resistance *vat(F)*. These genes help *A. veronii* to be resistant towards a number of antibiotic drugs such as phenicol, tetracyclines, sulfamethoxazole, beta-lactam class and macrolides etc. (Tekedar *et al.*, 2020). *Aeromonas veronii* strains exhibit resistance to drugs that were often used in aquaculture, including penicillin G, chloramphenicol, levofloxacin, trimethoprim, oxacillin, tetracycline, norfloxacin, sulfonamide, ofloxacin, cefazolin, and ciprofloxacin but found to be susceptible to cefoperazone, cephradine, cefuroxime, cefoxitin, chloramphenicol, cefepime, cefotaxime, clarithromycin, gentamycin, streptomycin, tobramycin (Sun *et al.*, 2016). Tetracycline resistance is shown by *A. veronii* isolated from catfish (Nawaz *et al.*, 2006). Penicillin resistance is shown by an extract of *A. veronii* from moribund freshwater ornamental fish (Sreedharan *et al.*, 2013). Therefore, antibiotics are no longer a good option to treat bacterial diseases in aquaculture system because of the emergence of antibiotic resistance of pathogens. Probiotics, vaccines, and phytochemicals are a few non-antibiotic strategies that can be used in response to the rising antibiotic resistance among *A. veronii* strains.

8. PROBIOTICS

It is generally recognized that probiotics have the ability to influence aquatic animals' innate immune systems (Figure 2). Probiotics are living bacteria that, when taken in the right quantities,

can improve the immune and digestive systems, increase viability, promote development, and benefit the host (Munir *et al.*, 2016). To reach the intestinal tract undamaged and continue to be alive, the optimal probiotic should be resistant to acid and bile salts and nonpathogenic (Boyle *et al.*, 2006). Many endogenic bacteria, including *Lactobacillus*, *Bacillus* (*B. subtilis* and *B. velezensis*), and yeast have gained attention as prospective probiotic contenders in the aquaculture industries in recent years due to their high level of antagonistic activity, availability, and exo-enzyme synthesis (Banerjee & Ray, 2017). As feed additives, *Lactobacillus*, *Bacillus* and yeast can colonise the digestive tract and maintain the microenvironmental balance in aquatic animals, stimulate development, enhance immunity, and increase resistance towards diseases (Zhang *et al.*, 2022; Brar *et al.*, 2023). *Bacillus velezensis* is a heterotypic synonym of *B. amyloliquefaciens*. *B. velezensis* combats *A. veronii* by secreting compounds like surfactins, bacillomycin, amylocyclin and fengicins (Fan *et al.*, 2017). In a study by Yang *et al.* (2022), *B. subtilis* CK3 was found to be able to inhibit the growth of *A. veronii* in crayfish (Yang *et al.*, 2022). Fish are benefited from the probiotic effects of LAB like *Pediococcus pentosaceus*, *Lactobacillus paraplantarum*, *Lactococcus lactis*, and *Enterococcus faecalis* (Martinez *et al.*, 2017; Meidong *et al.*, 2021; Heo *et al.*, 2013; Banos *et al.*, 2019). In order to increase probiotics' effectiveness in various animals (species and sizes) and environmental conditions, additional research should be conducted to identify the most suitable procedures and dosage levels (Di *et al.*, 2019). In order to avoid partially suppressing the immune responses, Farias *et al.* (2016) cautioned against feeding too many probiotics (Farias *et al.*, 2016). There are still gaps in our understanding of the fundamental mechanisms underlying the probiotics' beneficial and potentially harmful effects (Amenyogbe *et al.*, 2020). It is important to consider the dangers associated with probiotics producing antibiotics and transmission of antibiotic resistance genes to pathogenic bacteria (Martínez Cruz *et al.*, 2012).

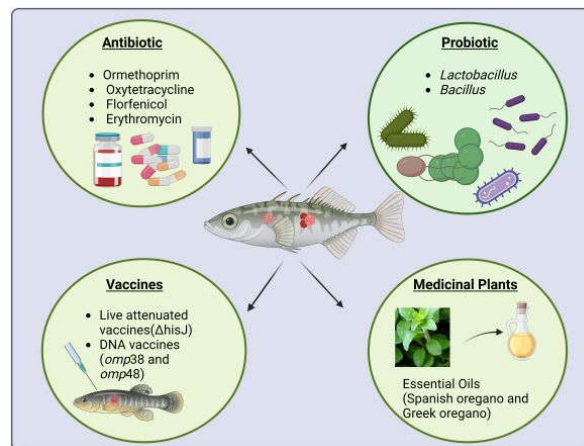


Figure 2: Different types of control measures used in aquaculture against *A. veronii*

Control strategies for bacterial infections caused by *Aeromonas* cover antibiotics, probiotics, vaccines and Herbal extracts.

9. VACCINES

It is generally known that vaccinations reduce the need for antibiotics and other antimicrobials and are a sustainable method of preventing serious fish infections. Currently, a variety of vaccinations, including subunit vaccines, DNA vaccines, live attenuated vaccines (LAVs), and formalin-killed vaccines are frequently used to prevent *A. veronii* infections (Huang *et al.*, 2014; Kong *et al.*, 2019). Vaccines have several benefits, including improved immunity, lower costs, reduced toxicity and drug side effects (Wu *et al.*, 2012). The effectiveness of commercial inactivated vaccines against MAS for some freshwater fish species depends entirely on the particular strain of the pathogenic bacteria (Ma *et al.*, 2019). Despite being approved in North America and Chile, live attenuated vaccines for catfish and salmonids are currently not commercially viable in other nations due to a lack of regulations governing their secure use in aquaculture (Adams, 2019). Zhang *et al.* (2020) found a live attenuated vaccine called " Δ hisJ" which is ideal for use in the development of a safe and effective immunisation against *A. veronii* infection in loach aquaculture (Zhang *et al.*, 2020). OMPs, or outer membrane proteins, (OmpF, OmpC, OmpR, OmpTS, Omp38, OmpA1, Omp48, OmpW, Tdr, 46 kDa maltoporin, and S-layer protein) are highly

immunogenic bacterial components. These OMPs are targeted by subunit vaccines for MAS because they have exposed cell epitopes, found at the host-bacterial interface and provide cross-protection due to their conserved antigenic determinants (Abdelhamed *et al.*, 2016). Iron-related protein A0KIY3, ATPase protein, fimbrial proteins Fim and FimMrfG, G-protein coupled receptor—GPR18, proaerolysin and hemolysin co-regulated protein—Hcp are additional antigenic proteins that could be used as recombinant protein vaccines (Pridgeon & Klesius, 2013; Zhang *et al.*, 2013; Guo *et al.*, 2018). Because subunit vaccines lack the components necessary to stimulate the immune system as effectively as a whole-cell vaccine, their capacity to elicit a potent immune response is constrained. Subunit vaccines are additionally expensive and unaffordable to scale up in developing nations due to the production and purification process (Dien *et al.*, 2023). By simulating the natural route of infection, DNA vaccination can produce protection against intracellular pathogens. DNA vaccination is a type of genetic immunisation that uses a gene or genes encoding protective antigens (Dien *et al.*, 2023). DNA vaccines using *Omp38* and *Omp48* genes show increased immune response in spotted sandbass (Vazquez-Juarez *et al.*, 2005). *OmpA1* and lactic acid bacteria can combine to create a dietary vaccine for carp against *A.*

veronii infection (Zhang *et al.*, 2018). The oral administration of the recombinant *L. casei* vaccine can successfully generate fish to establish cellular and humoral immunity that can colonize the intestine (Chen *et al.*, 2022). Furthermore, using DNA vaccine raises the additional safety concern of the link between intramuscular DNA vaccine administration and the onset of myositis, which needs to be clarified in further research (Dien *et al.*, 2023).

10. PHYTOCHEMICALS

To combat serious infections brought on by *Aeromonas* spp., botanicals and essential oils have also been utilised. More and more people are becoming interested in the antibacterial properties of natural materials and products, like the essential oils (EOs) obtained from medicinal aromatic plants (MAPs) (Cunha *et al.*, 2018). Because of their strong insecticidal, antiviral, antibacterial, and antifungal properties along with significant antioxidant activity, EOs stick out as effective alternative natural agents in fish aquaculture and animal husbandry for the prevention or treatment of a variety of infectious disorders (Elabd *et al.*, 2017). To reduce bacterial infections during fish rearing, EOs have been added to fish feed with encouraging results (Awad & Awaad, 2017). Few studies have examined the effectiveness of EOs and each of its separate elements against *A. veronii* to date (Anastasiou *et al.*, 2019). The EOs with the highest phenolic/carvacrol content are the most potent ones against *A. veronii*. More specifically, the EOs from savoury, Greek, and Spanish oregano are the most effective growth inhibitors of this particular fish illness (Mandalakis *et al.*, 2021). These pure EOs' increased effectiveness is most likely brought on by their higher concentrations of powerful chemicals including carvacrol (32.8% to 72.0%), γ -terpinene (5.3 to 34.0%), and p-cymene (6.5 to 11.9%). The most lethal EO is pure Spain oregano, which has a carvacrol content of 42.0%, not Greek oregano, which has a carvacrol content of 72.0%. So, it is evident that these EOs' ability to kill bacteria may be significantly influenced by other minor components and how they interact with the major components (Anastasiou *et al.*, 2019). It has been demonstrated that the Lamiaceae family members, especially MAPs rich in

carvacrol, have beneficial antibacterial properties (Cunha *et al.*, 2018). By affecting bacterial cell shape and deforming organelles, EOs primarily penetrate and operate on bacterial membrane and cytoplasm to impede their action mechanisms (Calo *et al.*, 2015). A variety of additional common vegetables, beans, and conventional herbs have been investigated for use in aquaculture as immunostimulants and antipathogens to manage *Aeromonas* infections in aquaculture including *Ocimum basilicum*, *Ocimum sanctum*, *Moringa oleifera*, *Mucuna pruriens*, *Aegle marmelos*, *Cynodon dactylon*, *Urtica dioica* and *Adrographis paniculata* (Kaleeswaran *et al.*, 2011; Das *et al.*, 2015). Combinations of various plant-based ingredients have demonstrated the potential to treat MAS in addition to the use of a single phytobiotic. In comparison to the group not given any medicinal herbs, the combination of huangqi root extract (*Astragalus membranaceus*) and Japanese honeysuckle flower extract (*Lonicera japonica*) reduced the mortality rate of Nile tilapia by 55% when administered orally (Ardó *et al.*, 2008). New phytochemical plants and novel combinations of probiotic- and plant-based products should be discovered through further research, which will also aim to improve the extraction process and standardise feed formulation, dose, and duration.

11. CONCLUSION

Globally widespread pathogen *Aeromonas veronii* has been one of the most significant bacteria influencing aquaculture and infects both humans and animals, causing heavy economic losses. The threat of an *A. veronii* infection in freshwater fish is constant to a number of fishes because they are surrounded by an *Aeromonas* environment. *Aeromonas veronii* may be hazardous to public health, particularly for individuals with weak immunity and those who are frequently exposed to infected fish. The control of *A. veronii* infection relies heavily on antibiotics. The improper use of antibiotics leads to the development of antibiotic-resistant genes in *A. veronii*. As a result, this pathogen is no longer susceptible to a number of antibiotics. Therefore, we ought to choose non-antibiotic tactics like probiotics, vaccinations, and phytochemicals. However, these techniques still

need to be studied further to increase their effectiveness and to get rid of any potential side effects. This review also omits some non-antibiotic strategies based on bacteriophage, omics, and nanobubble.

Acknowledgement- Authors duly acknowledge Central University of Himachal Pradesh for providing facilities to carry out the work.

Ethical Approval- 'Not applicable'

Consent to participate- Taken

Consent to Publish- Yes

Availability of data and materials- 'Not applicable'

Statements & Declarations:

Funding- "The authors declare that no funds, grants, or other support were received during the preparation of this manuscript."

Competing Interest: The authors declare no completing interest

Authors Contribution: RK- Conceptualization, AC, KT - Original draft preparation, KT & AKS- Reviewing and editing, BB, KC, DS, AS, SK, DM helped to draft the final manuscript, All authors have read and agreed to the published version of the manuscript.

REFERENCES

- Abd El Latif, A. M., Elabd, H., Amin, A., Eldeen, A. I., & Shaheen, A. A. (2019). High mortalities caused by *Aeromonas veronii*: identification, pathogenicity, and histopathological studies in *Oreochromis niloticus*. *Aquaculture International*, 27(6), 1725-1737.
- Abrami, L., Fivaz, M., Glauser, P. E., Sugimoto, N., Zurzolo, C., & van der Goot, F. G. (2003). Sensitivity of polarized epithelial cells to the pore-forming toxin aerolysin. *Infection and immunity*, 71(2), 739-746.
- Abdelhamed, H., Nho, S. W., Turaga, G., Banes, M. M., Karsi, A., & Lawrence, M. L. (2016). Protective efficacy of four recombinant fimbrial proteins of virulent *Aeromonas hydrophila* strain ML09-119 in channel catfish. *Veterinary Microbiology*, 197, 8-14.
- Adams, A. (2019). Progress, challenges and opportunities in fish vaccine development. *Fish & Shellfish Immunology*, 90, 210-214.
- Aguilera-Arreola, M. G., Hernandez-Rodriguez, C., Zuniga, G., Figueras, M. J., Garduño, R. A., & Castro-Escarpulli, G. (2007). Virulence potential and genetic diversity of *Aeromonas caviae*, *Aeromonas veronii*, and *Aeromonas hydrophila* clinical isolates from Mexico and Spain: a comparative study. *Canadian journal of microbiology*, 53(7), 877-887.
- Ahmad, A., Abdullah, S. R. S., Hasan, H. A., Othman, A. R., & Ismail, N. I. (2021). Aquaculture industry: Supply and demand, best practices, effluent and its current issues and treatment technology. *Journal of Environmental Management*, 287, 112271.
- Amal, M. N. A., Koh, C. B., Nurliyana, M., Suhaiba, M., Nor-Amalina, Z., Santha, S., & Zamri-Saad, M. (2018). A case of natural co-infection of Tilapia Lake Virus and *Aeromonas veronii* in a Malaysian red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) farm experiencing high mortality. *Aquaculture*, 485, 12-16.
- Amenyogbe, E., Chen, G., Wang, Z., Huang, J., Huang, B., & Li, H. (2020). The exploitation of probiotics, prebiotics and synbiotics in aquaculture: Present study, limitations and future directions.: A review. *Aquaculture International*, 28, 1017-1041.
- Anastasiou, T. I., Mandalakis, M., Krigas, N., Vézignol, T., Lazari, D., Katharios, P., & Antonopoulou, E. (2019). Comparative evaluation of essential oils from medicinal-aromatic plants of Greece: Chemical composition, antioxidant capacity and antimicrobial activity against bacterial fish pathogens. *Molecules*, 25(1), 148.
- Ardó, L., Yin, G., Xu, P., Váradi, L., Szigeti, G., Jeney, Z., & Jeney, G. (2008). Chinese herbs (*Astragalus membranaceus* and

- Lonicera japonica*) and boron enhance the non-specific immune response of Nile tilapia (*Oreochromis niloticus*) and resistance against *Aeromonas hydrophila*. *Aquaculture*, 275(1-4), 26-33.
- Awad, E., & Awaad, A. (2017). Role of medicinal plants on growth performance and immune status in fish. *Fish & shellfish immunology*, 67, 40-54.
- Bacharach, E., Mishra, N., Briese, T., Zody, M. C., Kembou Tsofack, J. E., Zamostiano, R., & Lipkin, W. I. (2016). Characterization of a novel orthomyxo-like virus causing mass die-offs of tilapia. *MBio*, 7(2), e00431-16.
- Bae, J. S., Shim, S. H., Hwang, S. D., Kim, J. W., Park, D. W., & Park, C. I. (2013). Molecular cloning and expression analysis of interleukin (IL)-15 and IL-15 receptor α from rock bream, *Oplegnathus fasciatus*. *Fish & shellfish immunology*, 35(4), 1209-1215.
- Banerjee, G., & Ray, A. K. (2017). The advancement of probiotics research and its application in fish farming industries. *Research in veterinary science*, 115, 66-77.
- Baños, A., Ariza, J. J., Nuñez, C., Gil-Martínez, L., García-López, J. D., Martínez-Bueno, M., & Valdivia, E. (2019). Effects of *Enterococcus faecalis* UGRA10 and the enterocin AS-48 against the fish pathogen *Lactococcus garvieae*. *Studies in vitro and in vivo*. *Food microbiology*, 77, 69-77.
- Barger, P. C., Liles, M. R., Beck, B. H., & Newton, J. C. (2021). Differential production and secretion of potentially toxigenic extracellular proteins from hypervirulent *Aeromonas hydrophila* under biofilm and planktonic culture. *BMC microbiology*, 21(1), 1-15.
- Bates, J. M., Mittge, E., Kuhlman, J., Baden, K. N., Cheesman, S. E., & Guillemin, K. (2006). Distinct signals from the microbiota promote different aspects of zebrafish gut differentiation. *Developmental biology*, 297(2), 374-386.
- Boyle, R. J., Robins-Browne, R. M., & Tang, M. L. (2006). Probiotic use in clinical practice: what are the risks?. *The American journal of clinical nutrition*, 83(6), 1256-1264.
- Brar, B., Kumar, R., Sharma, D., Sharma, A. K., Thakur, K., Mahajan, D., & Kumar, R. (2023). Metagenomic analysis reveals diverse microbial community and potential functional roles in Baner rivulet, India. *Journal of Genetic Engineering and Biotechnology*, 21(1), 147.
- Cai, S. H., Wu, Z. H., Jian, J. C., Lu, Y. S., & Tang, J. F. (2012). Characterization of pathogenic *Aeromonas veronii* bv. *veronii* associated with ulcerative syndrome from Chinese longsnout catfish (*Leiocassis longirostris* Günther). *Brazilian Journal of Microbiology*, 43, 382-388.
- Calo, J. R., Crandall, P. G., O'Bryan, C. A., & Ricke, S. C. (2015). Essential oils as antimicrobials in food systems-A review. *Food control*, 54, 111-119.
- Chao, C. M., Gau, S. J., & Lai, C. C. (2012). *Aeromonas* genitourinary tract infection. *Journal of Infection*, 65(6), 573-575.
- Chen, C., Zu, S., Zhang, D., Zhao, Z., Ji, Y., Xi, H., & Gu, J. (2022). Oral vaccination with recombinant *Lactobacillus casei* expressing Aha1 fused with CTB as an adjuvant against *Aeromonas veronii* in common carp (*Cyprinus carpio*). *Microbial cell factories*, 21(1), 1-17.
- Chen, F., Sun, J., Han, Z., Yang, X., Xian, J. A., Lv, A., & Shi, H. (2019). Isolation, identification and characteristics of *Aeromonas veronii* from diseased crucian carp (*Carassius auratus gibelio*). *Frontiers in microbiology*, 10, 2742.
- Chen, P. L., Tsai, P. J., Chen, C. S., Lu, Y. C., Chen, H. M., Lee, N. Y., & Ko, W. C. (2015). *Aeromonas* stool isolates from individuals with or without diarrhea in southern Taiwan: Predominance of *Aeromonas veronii*. *Journal of Microbiology, Immunology and Infection*, 48(6), 618-624.
- Choudhary, M., Thakur, K., Brar, B., Kumar, S., Sharma, D., Kumar, R., & Mahajan, D. (2023). Status of fish assemblage structure in the Ganga and Indus riverine systems of the western Himalaya. *World Water Policy*, 9(3), 613-638.
- Da Cunha, J. A., Heinzmann, B. M., & Baldisserotto, B. (2018). The effects of essential oils and their major

- compounds on fish bacterial pathogens—a review. *Journal of applied microbiology*, 125(2), 328-344.
- Das, R., Raman, R. P., Saha, H., & Singh, R. (2015). Effect of *Ocimum sanctum* Linn.(Tulsi) extract on the immunity and survival of *Labeo rohita* (Hamilton) infected with *Aeromonas hydrophila*. *Aquaculture Research*, 46(5), 1111-1121.
- Defoirdt, T., Sorgeloos, P., & Bossier, P. (2011). Alternatives to antibiotics for the control of bacterial disease in aquaculture. *Current opinion in microbiology*, 14(3), 251-258.
- Di, J., Chu, Z., Zhang, S., Huang, J., Du, H., & Wei, Q. (2019). Evaluation of the potential probiotic *Bacillus subtilis* isolated from two ancient sturgeons on growth performance, serum immunity and disease resistance of *Acipenser dabryanus*. *Fish & Shellfish Immunology*, 93, 711-719.
- Dien, L. T., Ngo, T. P. H., Nguyen, T. V., Kayansamruaj, P., Salin, K. R., Mohan, C. V., Rodkhum, C., & Dong, H. T. (2023). Non-antibiotic approaches to combat motile *Aeromonas* infections in aquaculture: Current state of knowledge and future perspectives. *Reviews in Aquaculture*, 15(1), 333-366
- Dong, H. T., Nguyen, V. V., Le, H. D., Sangsuriya, P., Jitrakorn, S., Saksmerprom, V., & Rodkhum, C. (2015). Naturally concurrent infections of bacterial and viral pathogens in disease outbreaks in cultured Nile tilapia (*Oreochromis niloticus*) farms. *Aquaculture*, 448, 427-435.
- Du, Z., Wang, D., Zhang, N., Wang, T., Wu, Q., Xu, Z., Long, Y., Jiang, L., & Luo, W. (2020). A case of *Aeromonas veronii* infection in golden Taiwanese loach (*Paramisgurnus dabryanus*) at low water temperature. *Aquaculture Research*, 51(3), 1307-1312.
- Ehsan, R., Rahman, A., Paul, S. I., Ador, M. A. A., Haque, M. S., Akter, T., & Rahman, M. M. (2023). *Aeromonas veronii* isolated from climbing perch (*Anabas testudineus*) suffering from epizootic ulcerative syndrome (EUS). *Aquaculture and Fisheries*, 8(3), 288-295.
- Elabd, H., Wang, H. P., Shaheen, A., Yao, H., & Abbass, A. (2017). Anti-oxidative effects of some dietary supplements on Yellow perch (*Perca flavescens*) exposed to different physical stressors. *Aquaculture Reports*, 8, 21-30.
- El-Gohary, F. A., Zahran, E., Abd El-Gawad, E. A., El-Gohary, A. H., M. Abdelhamid, F., El-Mleeh, A., & Elsayed, M. M. (2020). Investigation of the prevalence, virulence genes, and antibiogram of motile *Aeromonads* isolated from Nile tilapia fish farms in Egypt and assessment of their water quality. *Animals*, 10(8), 1432.
- Elorza, A., Rodríguez-Lago, I., Martínez, P., Hidalgo, A., Aguirre, U., & Cabriada, J. L. (2020). Gastrointestinal infection with *Aeromonas*: incidence and relationship to inflammatory bowel disease. *Gastroenterología y Hepatología (English Edition)*, 43(10), 614-619.
- Fan, B., Blom, J., Klenk, H. P., & Borriess, R. (2017). *Bacillus amyloliquefaciens*, *Bacillus velezensis*, and *Bacillus siamensis* form an “operational group *B. amyloliquefaciens*” within the *B. subtilis* species complex. *Frontiers in microbiology*, 8, 22.
- FAO. Statistical Query Results. Aquaculture: Quantity (t) 2008-2017. World. Inland waters.637
Freshwater. Common carp. *Cyprinus carpio*
- Farias, T. H. V., Levy-Pereira, N., de Oliveira Alves, L., de Carla Dias, D., Tachibana, L., Pilarski, F., de Andrade Belo, M. A., & Ranzani-Paiva, M. J. T. (2016). Probiotic feeding improves the immunity of pacus, *Piaractus mesopotamicus*, during *Aeromonas hydrophila* infection. *Animal Feed Science and Technology*, 211, 137-144
- Fernández-Bravo, A., & Figueras, M. J. (2020). An update on the genus *Aeromonas*: Taxonomy, epidemiology, and pathogenicity. *Microorganisms*, 8(1), 129.
- Fu, M., Kuang, R., Wang, W., Yu, Y., Ai, T., Liu, X., & Yuan, G. (2021). Hepcidin protects yellow catfish (*Pelteobagrus fulvidraco*) against *Aeromonas veronii*-induced

- ascites disease by regulating iron metabolism. *Antibiotics*, 10(7), 848.
- Gao, S., Zhao, N., Amer, S., Qian, M., Lv, M., Zhao, Y., & Zhao, B. (2013). Protective efficacy of PLGA microspheres loaded with divalent DNA vaccine encoding the ompA gene of *Aeromonas veronii* and the hly gene of *Aeromonas hydrophila* in mice. *Vaccine*, 31(48), 5754-5759.
- Gao, X., Tong, S., Zhang, S., Chen, Q., Jiang, Z., Jiang, Q., & Zhang, X. (2020). *Aeromonas veronii* associated with red gill disease and its induced immune response in *Macrobrachium nipponense*. *Aquaculture Research*, 51(12), 5163-5174.
- Guo, Z., Lin, Y., Wang, X., Fu, Y., Lin, W., & Lin, X. (2018). The protective efficacy of four iron-related recombinant proteins and their single-walled carbon nanotube encapsulated counterparts against *Aeromonas hydrophila* infection in zebrafish. *Fish & Shellfish Immunology*, 82, 50-59.
- Gauthier, D. T. (2015). Bacterial zoonoses of fishes: a review and appraisal of evidence for linkages between fish and human infections. *The Veterinary Journal*, 203(1), 27-35.
- Gavín, R., Merino, S., Altarriba, M., Canals, R., Shaw, J. G., & Tomás, J. M. (2003). Lateral flagella are required for increased cell adherence, invasion and biofilm formation by *Aeromonas* spp. *FEMS microbiology letters*, 224(1), 77-83.
- Gowda, T. K., Reddy, V. R., Devleeschauwer, B., Zade, N. N., Chaudhari, S. P., Khan, W. A., & Patil, A. R. (2015). Isolation and seroprevalence of *Aeromonas* spp. among common food animals slaughtered in Nagpur, Central India. *Foodborne Pathogens and Disease*, 12(7), 626-630.
- Gui, J. F., Tang, Q., Li, Z., Liu, J., & De Silva, S. S. (Eds.). (2018). *Aquaculture in China: success stories and modern trends*. John Wiley & Sons.
- Han, B. A., Kramer, A. M., & Drake, J. M. (2016). Global patterns of zoonotic disease in mammals. *Trends in Parasitology*, 32(7), 565-577.
- Heo, W. S., Kim, Y. R., Kim, E. Y., Bai, S. C., & Kong, I. S. (2013). Effects of dietary probiotic, *Lactococcus lactis* subsp. *lactis* I2, supplementation on the growth and immune response of olive flounder (*Paralichthys olivaceus*). *Aquaculture*, 376, 20-24.
- Hoai, T. D., Trang, T. T., Van Tuyen, N., Giang, N. T. H., & Van Van, K. (2019). *Aeromonas veronii* caused disease and mortality in channel catfish in Vietnam. *Aquaculture*, 513, 734425.
- Hossain, S., De Silva, B. C. J., Dahanayake, P. S., & Heo, G. J. (2018). Characterization of virulence properties and multi-drug resistance profiles in motile *Aeromonas* spp. isolated from zebrafish (*Danio rerio*). *Letters in applied microbiology*, 67(6), 598-605.
- Huang, L. Y., Wang, K. Y., Xiao, D., Chen, D. F., Geng, Y., Wang, J., & Xiao, G. Y. (2014). Safety and immunogenicity of an oral DNA vaccine encoding Sip of *Streptococcus agalactiae* from Nile tilapia *Oreochromis niloticus* delivered by live attenuated *Salmonella typhimurium*. *Fish & Shellfish Immunology*, 38(1), 34-41.
- Kaleeswaran, B., Ilavenil, S., & Ravikumar, S. (2011). Dietary supplementation with *Cynodon dactylon* (L.) enhances innate immunity and disease resistance of Indian major carp, *Catla catla* (Ham.). *Fish & Shellfish Immunology*, 31(6), 953-962.
- Kaur, A., Holeyappa, S. A., Bansal, N., Kaur, V. I., & Tyagi, A. (2020). Ameliorative effect of turmeric supplementation in feed of *Labeo rohita* (Linn.) challenged with pathogenic *Aeromonas veronii*. *Aquaculture International*, 28(3), 1169-1182.
- Kemper, N. (2008). Veterinary antibiotics in the aquatic and terrestrial environment. *Ecological indicators*, 8(1), 1-13.
- Khalifa, A. Y., AlMalki, M. A., & Bekhet, G. M. (2021). Pathological and mortality findings associated with *Aeromonas hydrophila* from frog eggs in Al-Ahsa region of Saudi Arabia. *Aquaculture Research*, 52(3), 1227-1236.
- Klesius, P., & Wei Pridgeon, Y. (2011). Live Attenuated Bacterial Vaccines in

- Aquaculture. In *International Symposium on Tilapia in Aquaculture* (pp. 18-26).
- Kong, Y.-D., Kang, Y.-H., Tian, J.-X., Zhang, D.-X., Zhang, L., Tao, L.-T., Wu, T.-L., Li, Y., Wang, G.-Q., & Shan, X.-F. (2019). Oral immunization with recombinant *Lactobacillus casei* expressing flaB confers protection against *Aeromonas veronii* challenge in common carp, *Cyprinus carpio*. *Fish & Shellfish Immunology*, 87, 627-637.
- Lewbart, G. A. (2001, January). Bacteria and ornamental fish. In *Seminars in Avian and Exotic Pet Medicine* (Vol. 10, No. 1, pp. 48-56). WB Saunders.
- Li, F., Wang, W., Zhu, Z., Chen, A., Du, P., Wang, R., & Wang, D. (2015). Distribution, virulence-associated genes and antimicrobial resistance of *Aeromonas* isolates from diarrheal patients and water, China. *Journal of Infection*, 70(6), 600-608.
- Li, F., Wu, D., Gu, H. R., Yin, M., Ge, H. L., Liu, X. H., & Wang, Z. J. (2019). *Aeromonas hydrophila* and *Aeromonas veronii* cause motile *Aeromonas* septicemia in the cultured Chinese sucker, *Myxocyprinus asiaticus*. *Aquaculture Research*, 50(5), 1515-1526.
- Li, T., Raza, S. H. A., Yang, B., Sun, Y., Wang, G., Sun, W., & Shan, X. (2020). *Aeromonas veronii* infection in commercial freshwater fish: A potential threat to public health. *Animals*, 10(4), 608.
- Li, Y., & Cai, S.-H. (2011). Identification and pathogenicity of *Aeromonas sobria* on tail-rot disease in juvenile tilapia *Oreochromis niloticus*. *Current Microbiology*, 62, 623-627.
- Li, Z., Wang, X., Chen, C., Gao, J., & Lv, A. (2019). Transcriptome profiles in the spleen of African catfish (*Clarias gariepinus*) challenged with *Aeromonas veronii*. *Fish & Shellfish Immunology*, 86, 858-867.
- Liu, G., Li, J., Jiang, Z., Zhu, X., Gao, X., Jiang, Q., Wang, J., Wei, W., & Zhang, X. (2022). Pathogenicity of *Aeromonas veronii* causing mass mortalities of *Odontobutis potamophila* and its induced host immune response. *Fish & Shellfish Immunology*, 125, 180-189.
- Lü, A., Song, Y., Hu, X., Sun, J., Li, L., Pei, C., & Nie, G. (2016). *Aeromonas veronii*, associated with skin ulcerative syndrome, isolated from the goldfish (*Carassius auratus*) in China. *The Israeli Journal of Aquaculture-Bamidgeh*.
- Ma, J., Bruce, T. J., Jones, E. M., & Cain, K. D. (2019). A review of fish vaccine development strategies: Conventional methods and modern biotechnological approaches. *Microorganisms*, 7(11), 569.
- Mandalakis, M., Anastasiou, T. I., Martou, N., Keisaris, S., Greveniotis, V., Katharios, P., & Antonopoulou, E. (2021). Antibacterial Effects of Essential Oils of Seven Medicinal-Aromatic Plants Against the Fish Pathogen *Aeromonas veronii* bv. sobria: To Blend or Not to Blend?. *Molecules*, 26(9), 2731.
- Martinez, M. P., Gonzalez Pereyra, M. L., Pena, G. A., Poloni, V., Fernandez Juri, G., & Cavaglieri, L. R. (2017). *Pediococcus acidolactici* and *Pediococcus pentosaceus* isolated from a rainbow trout ecosystem have probiotic and ABF1 adsorbing/degrading abilities in vitro. *Food Additives & Contaminants: Part A*, 34(12), 2118-2130.
- Martínez Cruz, P., Ibáñez, A. L., Monroy Hermosillo, O. A., & Ramírez Saad, H. C. (2012). Use of probiotics in aquaculture. *International Scholarly Research Notices*, 2012.
- Meidong, R., Nakao, M., Sakai, K., & Tongpim, S. (2021). *Lactobacillus paraplantarum* L34b-2 derived from fermented food improves the growth, disease resistance and innate immunity in *Pangasius bocourti*. *Aquaculture*, 531, 735878.
- Miao, Y., Zhao, X., Adam, F. E. A., Xie, Q., Feng, H., Ding, J., Bai, X., Wang, J. & Yang, Z. (2023). Isolation and identification of *Aeromonas veronii* in sheep with fatal infection in China: A Case Report. *Microorganisms*, 11(2), 333.
- Mohan, B., Sethuraman, N., Verma, R., & Taneja, N. (2017). Speciation, clinical profile & antibiotic resistance in *Aeromonas* species isolated from cholera-like illnesses in a tertiary care hospital in

- north India. *The Indian Journal of Medical Research*, 146(Suppl 1), S53.
- Munir, M. B., Hashim, R., Chai, Y. H., Marsh, T. L., & Nor, S. A. M. (2016). Dietary prebiotics and probiotics influence growth performance, nutrient digestibility and the expression of immune regulatory genes in snakehead (*Channa striata*) fingerlings. *Aquaculture*, 460, 59-68.
- Nawaz, M., Sung, K., Khan, S. A., Khan, A. A., & Steele, R. (2006). Biochemical and molecular characterization of tetracycline-resistant *Aeromonas veronii* isolates from catfish. *Applied and environmental microbiology*, 72(10), 6461-6466.
- Parkos, J., & Wahl, D. (2014). Effects of common carp (*Cyprinus carpio*), an exotic fish, on Aquatic Ecosystems. Illinois Natural History Survey report of January/February 2000. University of Illinois Board of Trustees. Center for Aquatic Ecology.
- Pei, C., Song, H., Zhu, L., Qiao, D., Yan, Y., Li, L., Zhao, X., Zhang, J., Jiang, X., & Kong, X. (2021). Identification of *Aeromonas veronii* isolated from largemouth bass *Micropterus salmoides* and histopathological analysis. *Aquaculture*, 540, 736707.
- Preena, P. G., Dharmaratnam, A., Raj, N. S., Kumar, T. V. A., Raja, S. A., & Swaminathan, T. R. (2019). Antibiotic susceptibility pattern of bacteria isolated from freshwater ornamental fish, guppy showing bacterial disease. *Biologia*, 74(8), 1055-1062.
- Pridgeon, J. W., & Klesius, P. H. (2013). G-protein coupled receptor 18 (GPR18) in channel catfish: Expression analysis and efficacy as immunostimulant against *Aeromonas hydrophila* infection. *Fish & Shellfish Immunology*, 35(4), 1070-1078.
- Qin, G., Ai, X., Xu, J., & Yang, Y. (2023). Dual RNA-seq of spleens extracted from channel catfish infected with *Aeromonas veronii* reveals novel insights into host-pathogen interactions. *Ecotoxicology and Environmental Safety*, 252, 114609.
- Qin, S., Xiao, W., Zhou, C., Pu, Q., Deng, X., Lan, L., & Wu, M. (2022). *Pseudomonas aeruginosa*: pathogenesis, virulence factors, antibiotic resistance, interaction with host, technology advances and emerging therapeutics. *Signal Transduction and Targeted Therapy*, 7(1), 1-27.
- Rahman, M. M. (2015). Role of common carp (*Cyprinus carpio*) in aquaculture production systems. *Frontiers in Life Science*, 8(4), 399-410.
- Ran, C., Qin, C., Xie, M., Zhang, J., Li, J., Xie, Y., & Zhou, Z. (2018). *Aeromonas veronii* and aerolysin are important for the pathogenesis of motile aeromonad septicemia in cyprinid fish. *Environmental microbiology*, 20(9), 3442-3456.
- Saavedra, M. J., Guedes-Novais, S., Alves, A., Rema, P., Tacão, M., Correia, A., & Martínez-Murcia, A. (2004). Resistance to β -lactam antibiotics in *Aeromonas hydrophila* isolated from rainbow trout (*Onchorhynchus mykiss*). *International microbiology*, 7(3), 207-211.
- Sahoo, P. K., Swaminathan, T. R., Abraham, T. J., Kumar, R., Pattanayak, S., Mohapatra, A., & Jena, J. K. (2016). Detection of goldfish haematopoietic necrosis herpes virus (Cyprinid herpesvirus-2) with multi-drug resistant *Aeromonas hydrophila* infection in goldfish: First evidence of any viral disease outbreak in ornamental freshwater aquaculture farms in India. *Acta tropica*, 161, 8-17.
- Sen, K., & Rodgers, M. (2004). Distribution of six virulence factors in *Aeromonas* species isolated from US drinking water utilities: a PCR identification. *Journal of applied microbiology*, 97(5), 1077-1086.
- Shameena, S. S., Kumar, K., Kumar, S., Kumar, S., & Rathore, G. (2020). Virulence characteristics of *Aeromonas veronii* biovars isolated from infected freshwater goldfish (*Carassius auratus*). *Aquaculture*, 518, 734819.
- Silver, A. C., Williams, D., Faucher, J., Horneman, A. J., Gogarten, J. P., & Graf, J. (2011). Complex evolutionary history of the *Aeromonas veronii* group revealed by host interaction and DNA sequence data. *PloS one*, 6(2), e16751.

- Song, H., Yang, B., Kang, Y., & Cong, W. (2023). Critical roles of VipB protein on virulence and oxidative stress tolerance in *Aeromonas veronii*. *Journal of Fish Diseases*, 46(5), 487-497.
- Song, Y., Hu, X., Lü, A., Sun, J., Yiksung, Y., Pei, C., Zhang, C., & Li, L. (2017). Isolation and characterization of *Aeromonas veronii* from ornamental fish species in China. *Israeli Journal of Aquaculture-Bamidgeh*, 69(1).
- Sreedharan, K., Philip, R., & Singh, I. S. B. (2013). Characterization and virulence potential of phenotypically diverse *Aeromonas veronii* isolates recovered from moribund freshwater ornamental fishes of Kerala, India. *Antonie Van Leeuwenhoek*, 103(1), 53-67.
- Sreedharan, K., Philip, R., & Singh, I. B. (2011). Isolation and characterization of virulent *Aeromonas veronii* from ascitic fluid of oscar *Astronotus ocellatus* showing signs of infectious dropsy. *Diseases of aquatic organisms*, 94(1), 29-39.
- Sun, J., Zhang, X., Gao, X., Jiang, Q., Wen, Y., & Lin, L. (2016). Characterization of virulence properties of *Aeromonas veronii* isolated from diseased Gibel Carp (*Carassius gibelio*). *International Journal of Molecular Sciences*, 17(4), 496.
- Sung, H. H., Hwang, S. F., & Tasi, F. M. (2000). Responses of giant freshwater prawn (*Macrobrachium rosenbergii*) to challenge by two strains of *Aeromonas* spp. *Journal of invertebrate pathology*, 76(4), 278-284.
- Tekedar, H. C., Arick, M. A., Hsu, C.-Y., Thrash, A., Blom, J., Lawrence, M. L., & Abdelhamed, H. (2020). Identification of antimicrobial resistance determinants in *Aeromonas veronii* strain MS-17-88 recovered from channel catfish (*Ictalurus punctatus*). *Frontiers in Cellular and Infection Microbiology*, 10, 348.
- Thakur, K., Sharma, A., Sharma, D., Brar, B., Choudhary, K., Sharma, A. K., & Kumar, R. (2022). An insight into the interaction between *Argulus siamensis* and *Labeo rohita* offers future therapeutic strategy to combat argulosis. *Aquaculture International*, 31, 1607-1621.
- Thakur, K., Mahajan, D., Sharma, A. K., Patial, P., Kumar, S., & Kumar, R. (2025). Understanding fish assemblage structure using enviro assessment techniques in a Northwestern Himalayan reservoir of Beas River basin in Himachal Pradesh (HP), India. *Environmental Science and Pollution Research*, 1-18.
- Tsai, Y. H., Shen, S. H., Yang, T. Y., Chen, P. H., Huang, K. C., & Lee, M. S. (2015). Monomicrobial necrotizing fasciitis caused by *Aeromonas hydrophila* and *Klebsiella pneumoniae*. *Medical Principles and Practice*, 24(5), 416-423.
- Tyagi, A., Sharma, C., Srivastava, A., Kumar, B. N., Pathak, D., & Rai, S. (2022). Isolation, characterization and complete genome sequencing of fish pathogenic *Aeromonas veronii* from diseased *Labeo rohita*. *Aquaculture*, 553, 738085.
- Vazquez-Juarez, R. C., Gomez-Chiarri, M., Barrera-Saldaña, H., Hernandez-Saavedra, N., Dumas, S., & Ascencio, F. (2005). Evaluation of DNA vaccination of spotted sand bass (*Paralabrax maculatofasciatus*) with two major outer-membrane protein-encoding genes from *Aeromonas veronii*. *Fish & shellfish immunology*, 19(2), 153-163.
- Vila, J., Ruiz, J., Gallardo, F., Vargas, M., Soler, L., Figueras, M. J., & Gascon, J. (2003). *Aeromonas* spp. and traveler's diarrhea: clinical features and antimicrobial resistance. *Emerging infectious diseases*, 9(5), 552.
- Vivekanandhan, G., Savithamani, K., Hatha, A. A. M., & Lakshmanaperumalsamy, P. (2002). Antibiotic resistance of *Aeromonas hydrophila* isolated from marketed fish and prawn of South India. *International journal of food microbiology*, 76(1-2), 165-168.
- Wahli, T., Burr, S. E., Pugovkin, D., Mueller, O., & Frey, J. (2005). *Aeromonas sobria*, a causative agent of disease in farmed perch, *Perca fluviatilis* L. *Journal of fish diseases*, 28(3), 141-150.
- Wang, H., Gu, Y., Luo, G., & Cao, H. (2020). *Aeromonas veronii*, a potential pathogen

- p>of enteritis in snakehead fish
- Ophiocephalus argus*
- .
- The Israeli Journal of Aquaculture-Bamidgeh*
- , 72.
- Wu, C. J., Chen, P. L., Tang, H. J., Chen, H. M., Tseng, F. C., Shih, H. I., & Ko, W. C. (2014). Incidence of *Aeromonas bacteremia* in southern Taiwan: *Vibrio* and *Salmonella* bacteremia as comparators. *Journal of Microbiology, Immunology and Infection*, 47(2), 145-148.
- Wu, L., Jiang, Y. N., Tang, Q., Lin, H. X., Lu, C. P., & Yao, H. C. (2012). Development of an *Aeromonas hydrophila* recombinant extracellular protease vaccine. *Microbial pathogenesis*, 53(5-6), 183-188.
- Wu, Z., Qi, X., Qu, S., Ling, F., & Wang, G. (2021). Dietary supplementation of *Bacillus velezensis* B8 enhances immune response and resistance against *Aeromonas veronii* in grass carp. *Fish & Shellfish Immunology*, 115, 14-21.
- Yang, H., Zhang, M., Ji, T., Zhang, Y., Wei, W., & Liu, Q. (2022). *Bacillus subtilis* CK3 used as an aquatic additive probiotics enhanced the immune response of crayfish *Procambarus clarkii* against newly identified *Aeromonas veronii* pathogen. *Aquaculture Research*, 53(1), 255-264.
- Yang, Q., Zhao, M., Wang, K. Y., Wang, J., He, Y., Wang, E. L., & Lai, W. (2017). Multidrug-resistant *Aeromonas veronii* recovered from channel catfish (*Ictalurus punctatus*) in China: prevalence and mechanisms of fluoroquinolone resistance. *Microbial Drug Resistance*, 23(4), 473-479.
- Yu, J. H., & Park, S. W. (2008). Isolation of *Aeromonas sobria* from cultured mud loach, *Misgurnus mizolepis*. *Journal of fish pathology*, 21(1), 21-27.
- Yu, J.-H., Han, J.-J., Kim, H.-J., Kang, S.-G., & Park, S.-W. (2010). First report of *Aeromonas veronii* infection in farmed Israeli carp *Cyprinus carpio* in Korea. *Journal of Fish Pathology*, 23(2), 165-176.
- Yu, X. J., Xu, L. J., & Wu, F. X. (2020). China fishery statistical yearbook.
- Yue, K., & Shen, Y. (2022). An overview of disruptive technologies for aquaculture. *Aquaculture and Fisheries*, 7(2), 111-120.
- Yuwono, C., Wehrhahn, M. C., Liu, F., Riordan, S. M., & Zhang, L. (2021). The isolation of *Aeromonas* species and other common enteric bacterial pathogens from patients with gastroenteritis in an Australian population. *Microorganisms*, 9(7), 1440.
- Zhai, W., Wang, Q., Zhu, X., Jia, X., & Chen, L. (2023). Pathogenic infection and microbial composition of yellow catfish (*Pelteobagrus fulvidraco*) challenged by *Aeromonas veronii* and *Proteus mirabilis*. *Aquaculture and Fisheries*, 8(2), 166-173.
- Zhang, D. X., Kang, Y. H., Chen, L., Siddiqui, S. A., Wang, C. F., Qian, A. D., & Shan, X. F. (2018). Oral immunization with recombinant *Lactobacillus casei* expressing *OmpAI* confers protection against *Aeromonas veronii* challenge in common carp, *Cyprinus carpio*. *Fish & Shellfish Immunology*, 72, 552-563.
- Zhang, D., Wu, Z., Chen, X., Wang, H., & Guo, D. (2019). Effect of *Bacillus subtilis* on intestinal apoptosis of grass carp *Ctenopharyngodon idella* orally challenged with *Aeromonas hydrophila*. *Fisheries science*, 85(1), 187-197.
- Zhang, H. P., Chen, M. Y., Xu, Y. X., Xu, G. Y., Chen, J. R., Wang, Y. M., & Ma, H. X. (2020). An effective live attenuated vaccine against *Aeromonas veronii* infection in the loach (*Misgurnus anguillicaudatus*). *Fish & Shellfish Immunology*, 104, 269-278.
- Zhang, H., Xu, R., & Wang, Z. (2021). Contribution of oxidative stress to HIF-1-mediated profibrotic changes during the kidney damage. *Oxidative Medicine and Cellular Longevity*, 2021.
- Zhang, Y., Ji, T., Jiang, Y., Zheng, C., Yang, H., & Liu, Q. (2022). Long-term effects of three compound probiotics on water quality, growth performances, microbiota distributions and resistance to *Aeromonas veronii* in crucian carp *Carassius auratus gibelio*. *Fish & Shellfish Immunology*, 120, 233-241.

- Zhang, D., Pridgeon, J. W., & Klesius, P. H. (2013). Expression and activity of recombinant proaerolysin derived from *Aeromonas hydrophila* cultured from diseased channel catfish. *Veterinary Microbiology*, 165(3-4), 478-482.
- Zheng, W., Cao, H., & Yang, X. (2012). *Aeromonas veronii* infection in the cultured snakehead fish, *Ophiocephalus argus* (Cantor). *African Journal of Microbiology Research*, 6(44), 7218-7223.
- Zhu, M., Wang, X. R., Li, J., Li, G. Y., Liu, Z. P., & Mo, Z. L. (2016). Identification and virulence properties of *Aeromonas veronii* bv. *sobria* isolates causing an ulcerative syndrome of loach *Misgurnus anguillicaudatus*. *Journal of Fish Diseases*, 39(6), 777-781.
- Zhu, X., Qian, Q., Wu, C., Zhu, Y., Gao, X., Jiang, Q., Wang, J., Liu, G., & Zhang, X. (2022). of *Aeromonas veronii* Causing Mass Mortality of Largemouth Bass (*Micropterus salmoides*) and Its Induced Host Immune Response. *Microorganisms*, 10(11), 2198.
- Ziarati, M., Zorriehzahra, M. J., Hassantabar, F., Mehrabi, Z., Dhawan, M., Sharun, K., & Shamsi, S. (2022). Zoonotic diseases of fish and their prevention and control. *Veterinary Quarterly*, 1-42.
