

## Feeding Ecology of Fishes – A Mini Review

<sup>1</sup>Yahya Bakhtiyar\*, <sup>2</sup>Sinan Nissar, <sup>3</sup>Mohammad Yasir Arafat, and <sup>4</sup>Raheela Mushtaq

### Author's Affiliation:

<sup>1,2,3,4</sup>Fish Biology and Limnology Research Laboratory, Department of Zoology, University of Kashmir, Srinagar, Jammu and Kashmir 190006, India

### \*Corresponding author:

**Yahya Bakhtiyar,**

Fish Biology and Limnology Research Laboratory, Department of Zoology, University of Kashmir, Srinagar, Jammu and Kashmir 190006, India  
E-mail: [yahya.bakhtiyar@gmail.com](mailto:yahya.bakhtiyar@gmail.com)

### Article Info:

Received on 22.02.2023

Revised on 23.03.2023

Approved on 19.05.2023

Accepted on 25.04.2023

Published on 16.06.2023

### ABSTRACT:

Fishes constitute more than half of the vertebrates, inhabiting almost every aquatic habitat of the world and form an inexpensive source of vital nutrients. The depletion of fishery stocks as a result of anthropogenic interferences is evident worldwide and mandates an urgent need to address the cause before rate of loss exceeds the limit. This makes the assessment of various aspects of fish, especially their feeding ecology quite imperative for developing conservation and management strategies. Food forms a vital factor for fishes required chiefly for growth and reproduction in fishes and its qualitative or quantitative variation is influenced by various biotic and abiotic factors. Amongst the various attributes influencing fishes, food and feeding constitute a vital aspect that impacts the growth and general wellbeing of the fish. Feeding ecology of a fish is directly linked to its population dynamics and helps in understanding of various aspects of fish like habitat preference, competition, prey selectivity, energy transfer etc. Under this backdrop the current review was drafted to acquire knowledge about the three major aspects of feeding ecology of fish i.e., analysis of the gut contents, feeding biology and prey selectivity. The analysis of gut contents helps to identify the prey diversity and abundance consumed by a fish. Feeding biology helps to evaluate the acceptability of the prey by depicting the correlation between biology of the fish and its food preferences. Prey selectivity deals with the evaluation of prey choice depicting the reason behind choice of one prey over the other.

**Keywords:** Conservation, Gut content analysis, Prey selectivity, Abiotic factors

**How to cite this article:** Bakhtiyar Y., Nissar S., Arafat M.Y., and Mushtaq R. (2023). Feeding Ecology of Fishes – A Mini Review. *Bulletin of Pure and Applied Sciences-Zoology*, 42A (1), 190-213.

## INTRODUCTION

With approximately 35,100 species, fishes form more than half of the vertebrates, inhabiting almost every aquatic habitat of the world (Froese and Pauly, 2023). Fish acts as an inexpensive source of vital nutrients, especially proteins, lipids, minerals, and vitamins (A and D), as such termed as the rich food for the impoverished (Sujatha *et al.*, 2013; Mohanty *et al.*, 2019; Ali *et al.*, 2020). Overfishing, alongside other anthropogenic interferences (climate change, pollution, introduction of new species and encroachments) has led to a decrease in the fish stocks (Jackson *et al.*, 2001), eventually leading to the extinction of many fish species (e.g., Pauly *et al.*, 2002; Scheffer *et al.*, 2005; Heithaus *et al.*, 2008). Such a scenario mandates critical action to prevent biodiversity loss, before the loss exceeds the limits (Rockstrom *et al.*, 2009). The biodiversity loss also mandates extensive research in various fields (feeding, population dynamics, invasive species, etc.) to form baseline data for various species to facilitate their better conservation.

A wide variety exists in the body forms of the fish, ranging from the small *Paedocypris progenetica* (8 mm total length) to the large *Rhincodon typus* (12 m total length), with the latter reaching around 15000 kg (Helfman *et al.*, 2009; Froese and Pauly, 2011). The vast difference in size is accompanied by variety of feeding habits, as fishes are found practically at every trophic level, from decomposers to herbivores to tertiary predators (Gerking, 1994; Wootton, 1998). Variation is further found with the generalists having a wider food preference range in comparison to the others that rely on specific food items like fins or scales (Nelson, 2006; Winemiller *et al.*, 2008). Besides the feeding mode in fishes depends on various morphological features, especially the shape of the body, length of the gut, gill rakers, size of the mouth, etc. (Cailliet *et al.*, 1996; Wootton, 1998). The type of food consumed by a fish also impacts its behavioural traits (Jobling, 1995).

Of all the attributes influencing the physiology, ecology, and general biology of the fishes, food

and feeding constitute an extremely cardinal aspect influencing the survival and general health of the fish. Since almost all the fishes are characterized by typical type II survivorship curves with heavy mortality in the larval stages (Begon *et al.*, 1990), therefore, the availability of the right type of food at right time and that too in the right concentration needs to be deliberated upon with assertion, as fishes are diverse with quite different feeding habits. Feeding ecology primarily helps to explore the strategy an animal opts for the optimum foraging of its preferred food and how a fish selectively ingests some organisms preferred over others, besides analyzing the multiple modes of feeding adaptations that comply with their morphological, sensory, and physiological responses to the type and abundance of food in the habitat (Wootton, 1998). Food and trophic level preferences play a critical role in the growth and development of fishes, due to their complex life history and often shift during the course of their life. Feed also forms the main input of aquaculture, as such the genre of feeding ecology has gained quite an impetus in recent years. Various workers (Helawell and Abel, 1971; Hyslop, 1980) have proposed that studies on the diet composition play a useful role in fishery science to comprehend trophic interactions, whereas the studies on the food and feeding habits of fishes help in determining their niche in the aquatic ecosystem, the preferred food items by the same further facilitate to determine the food spectrum overlapping with that of coexisting fishes (Yeon *et al.*, 1999). Further knowledge of an animal's dietary habits is a prerequisite to study its nutritional requirements, interactions with other organisms, and successful culture (Santos and Borges, 2001). The behaviour and population dynamics of a species are intricately linked with its feeding ecology (Braga *et al.*, 2012) which helps in understanding the subjects of habitat preference (Alimohammadi *et al.*, 2022; Adamczuk, 2022), resource partitioning (Ross, 1986; Guedes and Araújo, 2008; Alimohammadi *et al.*, 2022), predation (Martin *et al.*, 2005; Frid and Marliave, 2010), prey selection (Motta and Wilga, 2001), and energy transfer (Nakano and Murakami, 2001; Baxter *et al.*, 2004, 2005;

Rezende *et al.*, 2008). These ecological subjects are of great importance while developing conservation strategies for species and their ecosystems (Simpfendorfer *et al.*, 2011). The genre of feeding ecology is not a novel (e.g., Northcote, 1954), but emerged as a concept probably in the early 1980s (Cummis & Klug, 1979). This concept was refined in case of fishes by Gerking (1994) under the backdrop of optimal foraging theory in the context of physiological adaptation of fish.

## FEEDING ECOLOGY OF FISHES IN NATURE

Fishes are either herbivorous, carnivorous, or omnivorous and their body adapts morphologically, physiologically, and anatomically with their feeding (Wainwright, 1988, 1996; Yashpal, 2009). Alteration in several morphometric parameters has a direct relationship with feeding and body form is conventionally considered a strong attribute to successful feeding in fish (Bohórquez-Herrera *et al.*, 2015). Studies also reveal a functional relationship between gut microbes in a fish and the feeding habits (Bolnick *et al.*, 2014), sensory

organs, and digestive functions, that adjust to optimize the feeding process (Wilkens *et al.*, 2001; Scharnweber *et al.*, 2013). Environment invariably plays an important role in governing the food and feeding habits of different fish stocks and fish populations. The food and feeding in fishes is greatly influenced by many factors including temperature, light, salinity, fish size, activity and behaviour, appetite, feeding regime, starvation, stress, and type of food (Webb, 1978; Assan *et al.*, 2021). Nikolsky (1961) stressed that food supply is not only governed by the conditions for obtaining and utilizing food but also depends on many ecological conditions experienced during the feeding period such as temperature, illumination, winds, fluctuations in the water level, and changes in the feeding area. Many fishes exhibit great flexibility in their trophic ecology, for example, ontogenetic and seasonal changes in diet composition (Wootton, 1998; Costalago *et al.*, 2012; Fanelli *et al.*, 2022). The genre of the feeding ecology of fish could be studied under three main headings, gut content analysis, feeding biology, and prey selectivity as depicted in figure 1.

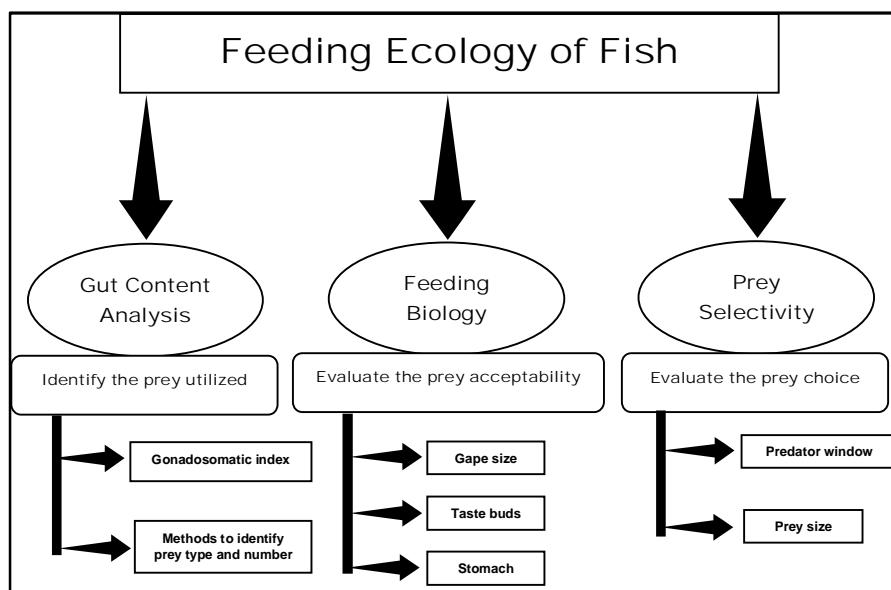


Figure 1: The various aspects studied under the genre of the feeding ecology of fish

## GUT CONTENT ANALYSIS

The knowledge about the diet of animals based on the analysis of gut contents is fundamental to the understanding of nutritional requirements and their interactions with other organisms (Windell and Bowen, 1978). Therefore, studies to evaluate the gut content try to identify and quantify the resources utilized by the species, providing information on those selected from the choices available in the environment (Williams, 1981; Tararam *et al.*, 1993). The process of gut content analysis requires the measurement of the length and weight of the fish specimen followed by dissecting its gut and preserving it in 5% formalin. This content is further analyzed using a binocular microscope. The gastrosomatic index of the fish is calculated using the formula given below:

$$\text{Gastrosomatic Index (GSI)} = \frac{\text{Weight of Stomach} \times 100}{\text{Weight of Fish}}$$

The contents in the fish gut are analyzed qualitatively and quantitatively as well (Hynes, 1950). There are three main approaches for the analysis of the fish gut i.e., bulk, numerical, and presence or absence, with the former comprising mass reconstruction, gravimetric, volumetric, point, and relative-fullness methods (Hyslop, 1980). The presence or absence method has the simplest approaches whereas the others are quite laborious. Many workers are of the view that stomach contents are based on prey consumed shortly before capture, but often it is observed that high percentage of stomachs are empty (Divita *et al.*, 1983; Brewer *et al.*, 1991; Bakhtiyar *et al.*, 2017) and therefore, these kinds of analysis represent the limited view of the diet in time and space (Pinnegar *et al.*, 2002; de la Mormiere *et al.*, 2003). A variety of indices, particularly the index of preponderance (IOP) (Natarajan and Jhingran, 1961), the index of relative importance (IRI) (Hyslop, 1980), the feeding index (FI) (Kawakami and Vazzoler, 1980) and the proposed and derived indices by Mohan and Sankaran (1988), Costello (1990) and Amundsen *et al.*, (1996). have been used for sampling of either undigested/poorly digested

gut contents to obtain information on the prey consumption (Pinkas, 1971; Manko, 2016; Mahesh *et al.*, 2018). Despite the statistical loopholes, these measures are frequently used due to easy computation and homogeneity, minimizing the error by enhancing the sample size (Carss, 1995). The studies have further been carried out by various workers to overcome the constraints met in gut analysis by improving and refining the methodology time by time. According to Windell and Bowen (1978), the selection of an appropriate technique will ultimately be determined by the investigation type, the presented hypothesis, or the nature of the food to be analyzed, though at times, equipment, time scale, or site restriction may take precedence. Several scientists have expressed their concern about the choice of analytic methods to observe the stomach contents of fish. Hyslop (1980) and Bowen (1983) proposed four main methods to study the gut content of fish, these are - numerical, weight, occurrence, and volumetric frequencies. The studies of the stomach contents of fishes suggest that one of the three methods becomes necessary to evaluate the abundance of the food items in the sample. Many types of conjugated data have been used to get the maximum possible information initiated from an inspection of the contents of the fish stomach. Amundsen and Sánchez-Hernández (2019) have provided a critical review of the various methods of analyzing stomach contents, suggesting the combination of the presence or absence method and relative fullness method for reliable results. Recent approaches in fish dietary studies involve the use of DNA barcoding of prey items (Kress *et al.*, 2015; Jakubaviciūtė *et al.*, 2017), however, it being a novel approach has some constraints yet to overcome. The principle involved for all of the mentioned methods is that food items should be counted or at least weighed or measured by their volume. Dietary descriptions of fish and other aquatic invertebrates are greatly influenced by the choice of method to quantify the relative importance of each prey to the diet. However, Braga (1999) and Lima-Junior (2000) proposed that the separation of food items for counting, weighing, or volumetric quantifying in an

individual way is impossible. Thus, recently a new method of analysis of fish stomach contents has been given by Lima-Junior and Goitein (2001). The most commonly used measures (numerical abundance, frequency of occurrence, and volume or weight measures) convey different types of information on feeding habits (Mac Donald and Green, 1983; Cortes, 1997, 1998; Amundsen and Sánchez-Hernández, 2019).

## FEEDING BIOLOGY

After acquiring information regarding the preferred food of a fish, one needs to look into the functional correlation between the biology and food preferences of the fish. The integration of biological evidence (based on food selectivity) with the mathematical indices could provide a better ecological hypothesis based on more scientific terms. Feeding ecology particularly shares its most interactive part with three aspects of fish biology i.e., physiological, morphological, and anatomical (Boglione *et al.*, 2003; Yúfera & Darias, 2007a, b). The procedure starts with the analysis of the morpho-ecology of mouth (Wainwright & Richard, 1995; Wainwright, 1996), which significantly defines the fish feeding habits during its ontogeny (Luczkovich *et al.*, 1995). Besides the feeding habits of fish changes with alterations in the surrounding environment and life stages. For instance, *Cyprinus carpio* exhibits periphytophagous feeding habit in rice fields but turns planktivorous in ponds (Saikia & Das, 2009). Similarly, *Labeo rohita* shows a carnivorous feeding habit in the fry stage and shifts to herbivorous diet upon reaching adulthood (Kamal, 1967). Similar reports on dietary shifts in response to prey availability and environmental variables have been demonstrated by Matsumoto and Kohda (2002) and Wang *et al.* (2019). Such flexibility in the diet breadth of a fish is facilitated by the plasticity of the mouth morphometry. Mouth gape acts as an entry point that limits the level of ingestion in a fish and has a strong correlation with the size of the prey a fish consumes at different stages of life (Knutsen & Tilseth, 1985; Mittelbach & Persson, 1998; Lukoschek & McCormick, 2001; Keppler *et al.*, 2015). Blaxter (1965) found gape size of the larvae of Atlantic herring as an

important morphological constraint in the early life of fish, also, according to Hyat (1979) and Dabrowski (1984), the gape size of the fish larvae is probably the most important morphological attribute that determines the behaviour and ecology of feeding. Being gape-limited predators, the larval fishes are initially constrained to small zooplankton prey, therefore, till the time they are gape-limited, the availability of small vulnerable zooplankton greatly influences the larval success (Zaret, 1980; Hansen and Wahl, 1981). Similarly, Dabrowski and Bardega (1984) studied the mouth size in the larvae of three cyprinid species to predict the size of food preferred by the fish at the initiation of exogenous feeding. They found a linear relationship between mouth size and the total length of fish, from the initial exogenous feeding stage up to 20-30mm. Ponton and Muller (1990) also found a linear relationship between gape size and the length of coregonous larvae. Unlike the adult planktivorous, larval fish are 'gape-limited' and as the larvae mature the gape size also starts increasing which thus allows them to take progressively larger prey (Lazzaro, 1987; Schael *et al.*, 1991; Bremigan and Stein, 1994). Hasan & McIntosh (1992) suggested the optimum particle food size for the carp fry, whereas Mookerji and Rao (1993) have reported a direct and linear increase in the mean size of the particle taken by the larvae of rohu (*Labeo rohita*) and singhi (*Heteropneustes fossilis*) as the age and the gape size increases. The feeding pattern (ram or suction) is an understated feature that can be inferred by the gape size of the fish (Wainwright & Richard, 1995). Two indices viz., Mouth area (MA) (Erzini *et al.* (1997) and Gape size (GS 90°) (Ponton & Müller, 1990) give us an estimation of the fish gape size. Morphologically, the vertical and horizontal width of the mouth and the head shape can be considered to explain the prey selectivity of various fishes.

Taste buds, a chemoreceptor or gustatory mechanoreceptor localized in the mouth, tongue, gills, and branchial cavity (Whitaker, 1971; Finger, 1997) play a vital role in the feeding mechanism of fish. These receptors are quite responsive to a wide array of substrates, especially the amino acids having a well-

coordinated system for diet selection (Oikeet *et al.*, 2007).

Immunohistochemical analysis (fluorescent dye assisted ligand specific induction followed by histomicrography) of taste buds could establish their role in food selectivity of fishes (Døving *et al.*, 2009). There are instances of food rejection in fishes even after ingestion into the mouth cavity (Bardach *et al.*, 1959; Gerhart *et al.*, 1991; Schulte and Bakus, 1992), indicating the involvement of receptors within the mouth cavity of the fish in the evaluation of food items (Kasumyan and Døving, 2003). The involvement of the gustatory sensory system as the final evaluator of feeding process is well-known scientifically and complemented by the anecdotes from fishermen, depicting the importance of taste properties of baits and feeds on food consumption, fishing success, and growth rate of fish (Jones, 1984; Takeda and Takii, 1992; Kasumyan, 1997). Fish possess more taste buds than other animals, with external as well as internal taste buds found in the former whereas only internal ones are found in the latter. In fish the taste buds are located on gill rakers, lips, oral cavity, oesophagus, pharynx, as well as on the body surfaces like barbels and fins (Ishimaru *et al.*, 2005). The external taste buds help in the detection and food selection of materials found in close proximity to the head but the oral taste buds play a vital role to determine the final food consumption (Kasumyan and Døving, 2003; Hansen and Reutter, 2004). The stomach forms the next major feature of the gut and accommodates recently ingested food items, as such could be used to characterize the food habits of a fish species. The first two coils of the guts are utilized for the same purpose in case of stomachless fish. The presence of the stomach and the length of the gut in relation to the body size (relative length of gut i.e., RLG) has been conventionally used as rough indicator of the feeding habit of a fish. Moreover, the analysis of gut microsections followed by enzymatic and bacterial assay analysis could provide more reliable information on the feeding habits of the fish (Bolnick *et al.*, 2014). Stomach content analysis of *Pterengraulis atherinoides* revealed the fish to be a specialized predator of Teleostei and juvenile Nanantia, with the diet varying with a

change in month and size (Krumme *et al.*, 2005). The gut contents analysis of *Argyrosomus japonicus* revealed mysid shrimps to be its most common prey item (for < 250mm total length fish), prawns were common in larger fish (301-450mm) and smaller fish were most common in the diet of fish larger than 500mm (Taylor *et al.*, 2006). The feeding ecology of *Delphinus delphis* revealed the diet to be dominated by smaller fish (especially *Notoscopelus kroyeri*), cephalopods, and crustaceans (Pusineri *et al.*, 2007).

### PREY SELECTIVITY

Prey selectivity pertains to the preference given by a fish species to prey over the other prey items, depending on several factors. It is quite a common feature of feeding in fish and has been the subject of a large number of studies. Selective foraging could be studied by comparing predator diets with the range of prey sizes available in the environment. Many workers have tried to explain selective foraging by designing theoretical, mechanistic models such as optimal foraging models (Stephens and Krebs, 1986) in manipulative experiments. It is observed that selective prey choice could arise either through passive processes (Scharf *et al.*, 1998) or through active choice by a predator (Sih and Christensen, 2001). It is further suggested that in aquatic environments, size is one of the major determinants of encounter rate or capture success because most populations of predators and prey are size-structured (Turesson *et al.*, 2002). Several reviews on the mechanism of prey selection by planktivorous fish (Eggers, 1977; Confer *et al.*, 1978; O'Brien *et al.*, 1979) have identified that differential visibility due to prey size, shape, color and motion, differential evasive ability among prey, and behavioral preference on the part of predator for certain prey, collectively contribute to prey selection. In case of piscivorous fishes, it is observed that when given a choice, piscivores feed upon the prey smaller than the maximum ingestible size (Webb, 1986; Wahl and Stein, 1988; Nilsson *et al.*, 1995 and Nilsson and Bronmark, 2000). Selectivity for a specific prey type at a given age must be an adaptive feature in larval fish to optimize energy intake (Greene, 1986).

Ontogenetic changes in prey selection patterns of larval fishes reveal that in nature larvae initiate feeding by engulfing small prey items and progressively go on selecting larger prey as they grow.

Fishes may be classified as stenophagous and euryphagous based on their diverse range of prey items (Oscoz *et al.*, 2005). Such classification is based on the concept of diet breadth, which explains the proportionality between ingested prey items and their availability in the environment. Despite the shortcomings of the gut analysis techniques, various diet breadth indices like, Levin's index (Hurlbert, 1978), and Manly's  $\alpha$  (Manly *et al.*, 1972; Chesson, 1978), etc are frequently used. Several other diet-based indices are associated with diet overlap and selectivity like, Ivlev's selectivity index (Ivlev, 1961) and Schoener's index (Schoener, 1968, 1970). Growth is one of the most important processes that determine the recruitment success during the early life history of fish (Crowder *et al.*, 1987). It is observed that growth in fish is strongly influenced by the availability of appropriate food items (Welker *et al.*, 1994). Most of the larval fish rely on small zooplankton as prey due to limited gape width (Bremigan and Stein, 1994; Devries *et al.*, 1998) and reduced visual acuity (Wahl *et al.*, 1993). Therefore, to understand processes at higher levels of organization, it becomes mandatory to understand the behavioural decisions made by individual predators and prey and the subsequent constraints that are imposed upon their behavioural strategies (Lima, 1998). Predators are usually selective foragers with respect to species and size of prey, where selectivity is defined as any difference in prey type composition and the predator diet compared to the composition of the available prey in the environment (Ivlev, 1961; Chesson, 1978).

While analyzing the ecology of 27 species of piscivorous freshwater fishes, Mittelbach and Persson (1998) showed that both the maximum and mean size of the prey eaten, increased with the predator size. During the culture of larvae also, it has been observed that prey size sequencing ensures optimal survival and growth rates (Tucker, 1998). Since size selectivity

is often interpreted as optimal foraging, in which, the predator encounters small prey but ignores it due to its low energetic content (Charnov *et al.*, 1976), however, it is further reported that differential encounter rate may be due to lower detectability of small prey (Sih and Christensen, 2001). Many workers have supported that minimum prey size has been referred to as predator's ability to detect its prey (Parma and Deriso, 1990; Lundvall *et al.*, 1999). Further, apart from prey size alone, Nilsson (2001) suggests that predator behaviour may seriously affect its prey, and size and density-dependent interactions between predators may be a major key to the understanding of predator-prey dynamics and community composition of aquatic habitat. Mechanisms, such as morphological constraints (Werner, 1974; Nilsson and Bronmark, 2000) or differences in spatial scales in combination with the swimming abilities of predator and prey (Christensen, 1996), may explain the maximum realized prey size a predator can take. A prey can be captured and eaten if the prey length: predator length ratio is within a specific range. This range has been referred to as the 'predation window' by Claessen *et al.* (2002). Earlier studies conducted by Alikunhi *et al.* (1955) suggested that the availability of live food organisms (zooplankton) of appropriate size and appropriate densities is probably one of the most critical factors affecting fry survival. Lubzens *et al.* (1984) have supported that during the first few days following yolk exhaustion, larvae of many carp species appear to feed either exclusively on zooplankton or feed very poorly when reared on other feeds, which are non-living and thus to overcome such problems an understanding of prey-predator interaction stands notable (Khadka and Rao, 1986). Gardner (1981) carried out experimental studies on the mechanisms of size selectivity in planktivorous fishes and indicated that the size selectivity results from the decisions made by the fish to ignore the small size class of *Daphnia*. Similar studies were carried out by Checkley Jr. (1982) in Atlantic Herring (*Clupea harengus*) larvae where it was observed that particles smaller than the largest acceptable size were consistently preferred by the predatory larvae. Rehage *et al.* (2005) conducted field and laboratory examinations to study the relationship between species foraging

behaviour and its impact by comparing the feeding behaviour of two non-invasive *Gambusia* species. Invasive *Gambusia* preferred *Daphnia*, avoided *Lirceus*, and consumed *Drosophila* in proportion to their availability. Larger fish consumed more prey items in their diets thus increasing diet breadth. Experimental studies by Colgan *et al.* (1986) on the feeding behaviour of largemouth bass (*Micropterus salmoides*) suggest that the natural diet of fry had significantly better net efficiency than the artificial diet of fry. The work of Khadka and Rao (1986) on the prey size selection by common carp larvae indicates that with increasing prey density the larvae selectively capture larger (more profitable) prey. Diet composition and prey preference of tench, perch, and roach were investigated by Giles *et al.* (1990). Experimental analysis of prey selection in the feeding of Largemouth Bass was carried out by Hambright (1991) who observed piscivorous fish to be the size-selective predators. Similar experimental studies were carried out by Gulbrandsen (1991) on Atlantic halibut larvae whereas Malhotra and Langer (1993) reported that fish larvae show preferential selection, which is determined by several factors viz. larval age, gape size, visibility, prey density, and prey digestibility. The works conducted by various other workers also confirm the factors influencing prey selectivity (Meng, 1993; Welker *et al.*, 1994; Wanzenbock, 1995; Mookerji and Rao, 1995; Liu and Uiblein, 1996; Bremigan and Stein, 1997; Limburg *et al.*, 1997 and Devries, *et al.*, 1998). Sirois and Dodson (2000) observed the critical period and growth-dependent survival

of an estuarine fish, rainbow smelt (*Osmerus mordax*). Experimental studies by Turesson *et al.* (2002) on the prey size selection in piscivorous pikeperch (*Stizostedion lucioperca*) indicate that pikeperch actively selects small-sized prey. Similar prey selectivity patterns were observed by Deudero and Morales-Nin (2001) in some planktivorous juvenile fishes. Predator-prey interactions with respect to prey size were investigated by Gill (2003) and Dorner and Wagner (2003). Fish size and prey availability and foraging behaviour in larval yellow perch (*Perca flavescens*) was investigated by Graeb *et al.* (2004). Similarly, Holzmann and Genin (2005) studied the mechanisms of selectivity in a nocturnal zooplanktivorous fish, *Apogon nannularis* and Cruz-Escalona *et al.* (2005) carried out observations on the feeding habits and trophic morphology of inshore lizardfish (*Synodus foetens*). Islam and Tanaka (2006) studied the changes in the diet of Japanese sea bass larvae with increasing size and changing ontogeny. Similar work on the ontogenetic shifts vis-à-vis interspecific diet similarity was undertaken by Nunn *et al.* (2007) whereas; selectivity patterns and food requirement of planktivorous alewife for two invasive predatory cladocerans were recorded by Pothoven *et al.* (2007). Prey selection patterns of common minke were recorded by Murase *et al.* (2007) while Lehtiniemi *et al.* (2007) studied the prey selectivity in three species of littoral fishes on nocturnal zooplankton assemblages. The feeding preferences of various commercially important fishes are compiled in table 1.

**Table 1: Depicts the feeding preferences of some economically important fish species**

Species	Feeding	Habitat	Reference
<i>Cyprinus carpio</i>	Omnivorous Main feed item: Crustaceans (Copepods) Alternatively feeds on algae	Freshwater (benthopelagic)	Sahtout <i>et al.</i> , 2018
	Feeds upon: Detritus, ostracods, macrophytes, zooplankton, insects, phytoplankton, and gastropods		Dadebo <i>et al.</i> , 2015
	Main prey item: Benthic insects, crustacea and detritus		Crivelli, 1981
	60 food items (22 Chlorophyceae, 12 Cyanobacteria, 10 Bacillariophyceae, and 16 Zooplankton) items found in gut content analysis		Saikia and Das, 2009



	Omnivorous Gut content analysis: 48.47% phytoplankton (mainly Bacillariophyceae and Chlorophyceae) and 51.53% zooplankton (mainly Rotifera)		Mondol <i>et al.</i> , 2013
<i>Ctenopharyngodon idella</i>	2.5 cm fish eat duckweed, <i>Spirodela</i> Adults prefer rooted plants	Freshwater (benthopelagic)	Lin, 1935
	Fish of size 11-15 mm TL may feed on Rotatoria, crustaceans, and sometimes chironomids and algae Fish of size 17-18 mm TL feed heavily on chironomids; reduced feeding on Rotatoria. Fish of size 30 mm feed exclusively on the microflora		Opuszynski, 1972
	Fish of size 17- 31 mm primarily consume benthic invertebrates Fish of size 32-86 mm chiefly consume periphyton and hydrilla and bank vegetation		Watkins <i>et al.</i> , 1981
<i>Hypophthalmichthys molitrix</i>	Silver carp diet was dominated by diatom ( <i>Cyclotella</i> spp.) followed by Chlorophyceae, Cyanophyceae, Crustacea, Dianophyceae, and Rotifera	Freshwater (benthopelagic)	Esmaeili and Johal, 2015
	Consumed zooplankton (Cladocera and Copepoda) and phytoplankton (Pyrrophyta, Chlorophyta, and Cyanophyta)		Spataru and Gophen, 1985
<i>Labeo rohita</i>	Fingerlings, preferred zooplankton ( <i>Arcella</i> and <i>Diffugia</i> , <i>Keratella</i> and <i>Brachionus</i> , <i>Daphnia</i> , and <i>Cyclops</i> ), and smaller algae ( <i>Cosmarium</i> , <i>Closterium</i> , <i>Euglena</i> , <i>Volvox</i> , algal spores and zygotes) while phytoplankton (green algae, diatoms, and blue-green algae) were avoided Adults showed a negative selection for zooplankton and a positive selection for green algae and diatoms ( <i>Ankistrodesmus</i> , <i>Spirogyra</i> , <i>Selenastrum</i> , <i>Scenedesmus</i> , <i>Tetraspora</i> , <i>Stephanodiscus</i> , <i>Diatoma</i> , <i>Synedra</i> , and <i>Nitzschia</i> ). Blue green algae were avoided	Freshwater (benthopelagic)	Khan and Siddiqui, 1973
	A herbi-omnivore in adult stages and an omnivore-planktophage in the early stages of life Early stage of life: algae, protozoans, rotifers, and Cladocerans were the most preferred food items Advanced stages: macrophytes, detritus, and sand/mud formed the major share of food.		Bakhtiyar <i>et al.</i> , 2017
<i>Catla catla</i>	Planktivorous (zooplankton feeder) Gut content analysis: Crustaceans (cladocerans and copepods) formed the main food item, followed by rotifers,	Freshwater	Lalit <i>et al.</i> , 2015

	Bacillariophyceae (diatoms), aquatic insects, Chlorophyceae, Myxophyceae, microvegetation, decayed and semi-decayed organic matter.		
<i>Oncorhynchus mykiss</i>	It is an active invertebrate predator Feeds preferentially on insect adults and larvae. Leptophlebiid may flies and dipteran pupae were the two most abundant food items in Blaylock Creek Representatives of seven insect orders, an isopod and a crayfish followed by Dipterans, Ephemeropterans, Trichopterans, and Hymenopterans were retrieved from the stomachs of the fish in Long Creek	Freshwater (benthopelagic)	Metcalf <i>et al.</i> , 1997
	Two most important food group: aquatic insects (89%) and fishes Intake of algae and plant material increased with size of fish Microcrustaceans (Cladocera and Copepoda) were also preferred by the smaller trout		Leonard and Leonard, 1949
<i>Salmo truttafario</i>	Most common food items: Brachycentridae, <i>Blepharocera</i> spp., Hydropsychidae, <i>Ephemerella</i> spp.	Freshwater (pelagic-neritic)	Khan <i>et al.</i> , 2021
	Main food items: Trichoptera, Ephemeroptera, Diptera, Plecoptera, Coleoptera, Odonata, Amphipoda, Hirudinea, Megaloptera, trout egg, plant seeds and terrestrial ants		Rasool <i>et al.</i> , 2012
<i>Salvelinus fontinalis</i>	Most common terrestrial prey: small Hymenopterans, Coleopterans, and Aphids Other frequently genera: Simuliids, Plecopterans, and water mites Low frequency of occurrence: Coleopterans (aquatic fraction), adult Trichopterans, Dipterans, and Psyllids (terrestrial fraction) Fishes and lizards were also observed from the stomach of few individuals	Freshwater (benthopelagic)	Horká <i>et al.</i> , 2017
<i>Brycinus nurse</i>	The relative importance (RI) indices of <i>Ceriodaphnia</i> sp., <i>Povillaadusta</i> eggs, <i>Ceriodaphnia</i> eggs and detritus were found to be higher in dry season as compared to the wet season	Freshwater (pelagic)	Saliu, 2002
<i>Oligosarcushepsetus</i>	<i>Cichlamonoculus</i> dominating in autumn Lepidoptera and Hymenoptera dominated in winter Hemiptera dominated the diet in summer	Freshwater (benthopelagic)	Araujo <i>et al.</i> , 2005
<i>Labeo niloticus</i>	Availability of food governed diet quality Variability of feeding attributed to breeding season and climate A bottom feeder, feeds on organic debris, soft and decaying vegetation, and small	Freshwater (benthopelagic)	El Moghraby and El Rehman, 1984

	organisms found in its habitat		
<i>Creagrutus brevipinnis</i>	A seasonal variation in the feeding activity of the fish Mainly feeds from 06:00 to 18:00 hrs	Freshwater (benthopelagic)	Roman-Valencia, 1998
<i>Chanos chanos</i>	Gut content analysis: Mainly single cellular green algae (Chlorophyceae) and filamentous blue-green algae (Cyanophyta) Small fraction: Diatoms, crustaceans (copepods, phyllopods, and nauplii), Ciliata, Dinoflagellata, and Rotatoria	Amphidromous (benthopelagic)	Lückstädt and Reiti, 2002
	Main food (19-27 cm): Phytoplankton (Cyanophyceae and Dinophyceae groups) <i>Navicula</i> and <i>Fragillaria</i> are preferred quite less		A'yun and Takarina, 2019
<i>Mugil cephalus</i>	Major food: Plant matter (diatoms, algae, and dinoflagellates) Animal origin food: Annelids, fish larvae, insect parts and crustaceans	Catadromous (benthopelagic)	Jamabo and Maduako, 2015
	Omnivorous feeder Highest amount: Sand and mud Basic food: Diatoms and algal matter		Lavanya <i>et al.</i> , 2018
	Bacillariophyceae as the most preferred food material Myxophyceae was given the second preference Dominant zooplankton in gut content Dinoflagellates, followed by copepods Besides parts of fish and shrimp also formed minor contents of stomach		Mondal <i>et al.</i> , 2015
<i>Etroplus suratensis</i>	Main food: Decayed organic matter (38.61%), filamentous algae (29.15%), and miscellaneous matter (8.04%)	Brackish (benthopelagic)	Joseph and Joseph, 1988
	Main components of gut: Aquatic macrophytes (33%) and filamentous algae (31%) Other components: Detritus and digested matter (12%), diatoms (11%), zooplankton (5%) and molluscs (1%)		Emmanuel <i>et al.</i> , 2019
	Major food items: Filamentous algae, detritus, aquatic plants and diatoms Other food items: Rotifers, insect larvae, Cladocerans, Copepods, and gastropods		Priya <i>et al.</i> , 2020
<i>Lates Latescalcarifer calcarifer</i>	Major items: Crustaceans (34%) and small fishes (22.0%) Other items: Mollusca (13%) and algae (9.5%)	Catadromous (demersal)	Panchakshari <i>et al.</i> , 2016
	Major components: Crustaceans (shrimps and crabs) and fish larvae Other components: Fish larvae, polychaetes and algae (Bacillariophyceae)		Krishna <i>et al.</i> , 2016
<i>Percalates colonorum</i>	Shows temporal, spatial and size class	Catadromous	Howell <i>et al.</i> ,

	variations in diet; Diet changed seasonally	demersal	2004
<i>Onchorhynchus tshawytscha</i>	<i>Engraulismordax</i> , <i>Sebastes</i> spp., <i>Clupea pallasii</i> and <i>Cancer magister</i> as main prey Seasonal and annual differences in the dominant prey items	Anadromous (benthopelagic)	Hunt <i>et al.</i> , 1999
<i>Salvelinus alpinus</i>	Feeding habit shows a dominance of zooplankton in late autumn and of chironomid larvae in winter and chironomid pupae in summers	Anadromous	Svenning <i>et al.</i> , 2007
<i>Lutjanus fulvivflamma</i>	Dawn and dusk as peak feeding hours Opportunistic feeders 46% specimens had empty stomach Brachyurans were the main prey item (48% Index of relative importance)	Marine/brackish (reef-associated)	Kamukuru and Mgaya, 2004
<i>Pseudaphritisurvilli</i>	Numerical and volumetric analysis of the gut contents of 698 specimens revealed the fish was a generalized omnivore, feeding on benthic animals	Catadromous (benthopelagic)	Horte and White, 1980
<i>Paralichthys orbignyanus</i>	Diet comprised of polychaetes	Oceanodromous (demersal)	Prisco <i>et al.</i> , 2001
<i>Dissostichus <u>eleginoides</u></i>	Wide variations in the diet with respect to the season, size, and depth of the ocean	Oceanodromous (pelagic-oceanic)	Arkhipkin <i>et al.</i> , 2003
<i>Synodus foetens</i>	<i>Loligo pealei</i> and <i>Upeneus parvus</i> being vital prey during nortes season <i>L. pealei</i> and <i>Engyophrysenta</i> were reported to be the main prey during rainy season	Marine/brackish (reef-associated)	Cruz-Escalona <i>et al.</i> , 2005.
<i>Trachurus trachurus</i>	Majority of empty stomachs in January and February and minimum during July and August	Oceanodromous (pelagic-neritic)	Santic <i>et al.</i> , 2005
<i>Pagrus pagrus</i>	Carnivorous feeding habit feeds primarily on decapods	Oceanodromous	Labropoulou <i>et al.</i> , 1999
<i>Oncopterus darwinii</i>	Diet comprised of small crustaceans	Marine (benthopelagic)	Prisco <i>et al.</i> , 2001.
<i>Notolabrus fucicola</i>	Major prey items are crabs, bivalves, and amphipods. 18 prey items from the stomach (microalgae, aquatic insects, terrestrial arthropods and plant matter)	Marine (reef-associated)	Denny and Schiel, 2001; Ortaz, 2001
<i>Rastrelliger kanagurta</i>	Mainly feed on pelagic crustaceans (copepods) represented 51.15% by weight. Fishes accounted 66.7% occurrence and 30.35% by weight Minor item from gut: Sand 38.8%by occurrence and scales 30.6%by occurrence	Oceanodromous (pelagic-neritic)	Nath <i>et al.</i> , 2015
<i>Sardachiliensis chiliensis</i>	Ichthyophagous and stenophagous fish Generalist predator Fish feed more during the autumn season. Main food items: <i>Pleuroncodes monodon</i> and <i>Engraulis ringens</i>	Oceanodromous (pelagic-neritic)	Pepe-Victoriano <i>et al.</i> , 2022
<i>Sardinella longiceps</i>	Mainly feeds on Diatoms Other food items: Dinoflagellates, copepods and fish eggs	Oceanodromous (pelagic-neritic)	Zaki <i>et al.</i> , 2021

### Conflict of interest

The authors have no conflicts of interest to declare that are relevant to the content of this article

### Acknowledgments

The authors are highly grateful to the Head, Department of Zoology, University of Kashmir for providing all the necessary facilities during the present study.

### REFERENCES

1. A'yun, Q., & Takarina, N. D. (2019). Food preference analysis of milkfish in Blanakan Ponds, Subang, West Java. *AIP Conference Proceedings*, 2168(1), 020083.
2. Adamczuk, M. (2022). The monitoring of diet and habitat preferences indicates competitive effect of exotic *Ictalurus nebulosus* on native fish under food-limited conditions. *Global Ecology and Conservation*, 34, e02060.
3. Ali, S. S. R., Abdhakir, E. S., Muthukkaruppan, R., Sheriff, M. A., & Ambasankar, K. (2020). Nutrient Composition of Some Marine Edible Fish Species from Kasimedu Fish Landing Centre, Chennai (TN), India. *International Journal of Biological Innovations*, 2(2), 165-173.
4. Alikunhi, K. H., Chaudhuri, H., & Ramachandran, V. (1955). On the mortality of carp fry in nursery ponds and the role of plankton in their survival and growth. *Indian Journal of Fisheries*, 2(2), 257-313.
5. Alimohammadi, M., Valinassab, T., Ramezani-Fard, E., & Ehteshami, F. (2022). Feed comparison and feeding ecology in five sympatric teleost species of the northern Oman Sea. *Iranian Journal of Fisheries Sciences*, 21(2), 463-479.
6. Amundsen, P. A., & Sánchez-Hernández, J. (2019). Feeding studies take guts—critical review and recommendations of methods for stomach contents analysis in fish. *Journal of Fish Biology*, 95(6), 1364-1373.
7. Amundsen, P. A., Gabler, H. M., & Staldvik, F. J. (1996). A new approach to graphical analysis of feeding strategy from stomach contents data—modification of the Costello (1990) method. *Journal of Fish Biology*, 48(4), 607-614.
8. Araújo, F. G., Andrade, C. C., Santos, R. N., Santos, A. F. G., & Santos, L. N. (2005). Spatial and seasonal changes in the diet of *Oligosarcus hepsetus* (Characiformes, Characidae) in a Brazilian reservoir. *Brazilian Journal of Biology*, 65, 1-8.
9. Arkhipkin, A., Brickle, P., & Laptikhovsky, V. (2003). Variation in the diet of the Patagonian tooth fish with size, depth and season around the Falkland Islands. *Journal of Fish Biology*, 63(2), 428-441.
10. Assan, D., Huang, Y., Mustapha, U. F., Addah, M. N., Li, G., & Chen, H. (2021). Fish feed intake, feeding behavior, and the physiological response of apelin to fasting and refeeding. *Frontiers in Endocrinology*, 12, 798903.
11. Bakhtiyar, Y., Langer, S., Karlopiya, S. K., & Chalotra, R. K. (2017). Studies on the feeding habits of *Labeorohita* (Ham.) from Gho-Manhasa fish ponds, Jammu, North India. *Journal of Ecophysiology and Occupational Health*, 17(1/2), 40-49.
12. Bardach, J. E., Winn, H. E., & Menzel, D. W. (1959). The role of the senses in the feeding of the nocturnal reef predators *Gymnothorax moringa* and *G. vicinus*. *Copeia*, 1959(2), 133-139. <https://doi.org/10.2307/1440065>
13. Baxter, C. V., Fausch, K. D., & Carl Saunders, W. (2005). Tangled webs: reciprocal flows of invertebrate prey link streams and riparian zones. *Freshwater Biology*, 50(2), 201-220.
14. Baxter, C. V., Fausch, K. D., Murakami, M., & Chapman, P. L. (2004). Fish invasion restructures stream and forest food webs by interrupting reciprocal preysubsidies. *Ecology*, 85(10), 2656-2663.
15. Begon, M., Harper, J.L., & Townsend, C.R. (1990). *Ecology: Individuals populations and communities* (2<sup>nd</sup> ed.). Oxford UK: Blackwell Scientific Publication.
16. Blaxter, J. H. S. (1965). The feeding of herring larvae and their ecology in relation to feeding. *CalCOFI Reports*, 10, 79-88.
17. Boglione, C., Giganti, M., Selmo, C., & Cataudella, S. (2003). Morphoecology in larval fin-fish: a new candidate species for aquaculture, *Diplodus puntazzo*

- (Sparidae). *Aquaculture International*, 11, 17-41.
18. Bohórquez-Herrera, J., Cruz-Escalona, V. H., Adams, D. C., & Peterson, M. S. (2015). Feeding ecomorphology of seven demersal marine fish species in the Mexican Pacific Ocean. *Environmental Biology of Fishes*, 98, 1459-1473.
19. Bolnick, D. I., Snowberg, L. K., Hirsch, P. E., Lauber, C. L., Knight, R., Caporaso, J. G., & Svanbäck, R. (2014). Individuals' diet diversity influences gut microbial diversity in two freshwater fish (threespine stickleback and Eurasian perch). *Ecology Letters*, 17(8), 979-987.
20. Bowen, S.H. (1983). Quantitative description of the diet. In L.A., Nielsen & D.L., Johnson (Eds.). *Fisheries techniques* (pp. 325-336). Maryland: American Fisheries Society.
21. Braga, F. M. de S. (1999). O grau de preferência alimentar: um método qualitativo e quantitativo para o estudo do conteúdo estomacal de peixes. *Acta Scientiarum. Biological Sciences*, 21(2), 291-295.
22. Braga, R. R., Bornatowski, H., & Vitule, J. R. S. (2012). Feeding ecology of fishes: an overview of worldwide publications. *Reviews in Fish Biology and Fisheries*, 22, 915-929.
23. Bremigan, M. T., & Stein, R. A. (1997). Experimental assessment of the influence of zooplankton size and density on gizzard shad recruitment. *Transactions of the American Fisheries Society*, 126(4), 622-637.
24. Bremigan, MT, & Stein, RA (1994). Gape-dependent larval foraging and zooplankton size: implications for fish recruitment across systems. *Canadian Journal of Fisheries and Aquatic Sciences*, 51 (4), 913-922.
25. Brewer, D. T., Blaber, S. J. M., & Salini, J. P. (1991). Predation on penaeid prawns by fishes in Albatross Bay, Gulf of Carpentaria. *Marine Biology*, 109(2), 231-240.
26. Cailliet, G.M., Love, M.S., & Ebeling, A.W. (1996). *Fishes: a field and laboratory manual on their structure, identification and natural history*. Belmont, CA: Wadsworth Publication.
27. Carss, D. (1995). Foraging behaviour and feeding ecology of the otter *Lutra lutra*: a selective review. *Hystrix, The Italian Journal of Mammalogy*, 7(1-2), 179-194.
28. Charnov, E. L., Orians, G. H., & Hyatt, K. (1976). Ecological implications of resource depression. *The American Naturalist*, 110(972), 247-259.
29. Checkley, D. M. (1982). Selective feeding by Atlantic herring (*Clupea harengus*) larvae on zooplankton in natural assemblages. *Marine Ecology Progress Series*, 9(3), 245-253.
30. Chesson, J. (1978). Measuring preference in selective predation. *Ecology*, 59(2), 211-215.
31. Christensen, B. (1996). Predator foraging capabilities and prey antipredator behaviours: pre-versus postcapture constraints on size-dependent predator-prey interactions. *Oikos*, 76(2), 368-380.
32. Claessen, D., Van Oss, C., de Roos, A. M., & Persson, L. (2002). The impact of size-dependent predation on population dynamics and individual life history. *Ecology*, 83(6), 1660-1675.
33. Colgan, P. W., Brown, J. A., & Orsatti, S. D. (1986). Role of diet and experience in the development of feeding behaviour in largemouth bass, *Micropterus salmoides*. *Journal of Fish Biology*, 28(2), 161-170.
34. Confer, J. L., Howick, G. L., Corzette, M. H., Kramer, S. L., Fitzgibbon, S., & Landesberg, R. (1978). Visual predation by planktivores. *Oikos*, 31, 27-37.
35. Cortés, E. (1997). A critical review of methods of studying fish feeding based on analysis of stomach contents: application to elasmobranch fishes. *Canadian Journal of Fisheries and Aquatic Sciences*, 54(3), 726-738.
36. Cortés, E. (1998). Methods of studying fish feeding: reply. *Canadian Journal of Fisheries and Aquatic Sciences*, 55(12), 2708.
37. Costalago, D., Navarro, J., Álvarez-Calleja, I., & Palomera, I. (2012). Ontogenetic and seasonal changes in the feeding habits and trophic levels of two small pelagic fish species. *Marine Ecology Progress Series*, 460, 169-181.
38. Costello, M. J. (1990). Predator feeding strategy and prey importance: a new graphical analysis. *Journal of Fish Biology*, 36(2), 261-263.

39. Crivelli, A. J. (1981). The biology of the common carp, *Cyprinus carpio* L. in the Camargue, southern France. *Journal of Fish Biology*, 18(3), 271-290.
40. Crowder, L. B., McDonald, M. E., & Rice, J. A. (1987). Understanding Recruitment of Lake Michigan Fishes: The Importance of Size-Based Interactions between Fish and Zooplankton. *Canadian Journal of Fisheries and Aquatic Sciences*, 44(S2), s141-s147.
41. Cruz-Escalona, H.V., Peterson, M.S., Davilla, C.L., & Zetina-Rejon, M. (2005). Feeding habits and trophic morphology of inshore lizardfish (*Synodus foetens*) on the central continental shelf off Veracruz, Gulf of Mexico. *Journal of Applied Ichthyology*, 21 (6), 525-530.
42. Cummins, K. W., & Klug, M. J. (1979). Feeding ecology of stream invertebrates. *Annual Review of Ecology and Systematics*, 10(1), 147-172.
43. Dabrowski, K. J. R. N. D. (1984). The feeding of fish larvae: present 'state of the art' and perspectives. *Reproduction Nutrition Development*, 24(6), 807-833.
44. Dabrowski, K., & Bardega, R. (1984). Mouth size and predicted food size preferences of larvae of three cyprinid fish species. *Aquaculture*, 40(1), 41-46.
45. Dadebo, E., Eyayu, A., Sorsa, S., & Tilahun, G. (2015). Food and feeding habits of the common carp (*Cyprinus carpio* L. 1758) (Pisces: Cyprinidae) in Lake Koka, Ethiopia. *Momona Ethiopian Journal of Science*, 7(1), 16-31.
46. de la Morinière, E. C., Pollux, B. J. A., Nagelkerken, I., Hemminga, M. A., Huiskes, A. H. L., & van der Velde, G. (2003). Ontogenetic dietary changes of coral reef fishes in the mangrove-seagrass-reef continuum: stable isotopes and gut-content analysis. *Marine Ecology Progress Series*, 246, 279-289.
47. Denny, C. M., & Schiel, D. R. (2001). Feeding ecology of the banded wrasse *Notolabrus fucicola* (Labridae) in southern New Zealand: prey items, seasonal differences, and ontogenetic variation. *New Zealand Journal of Marine and Freshwater Research*, 35(5), 925-933.
48. Deudero, S., & Morales-Nin, B. (2001). Prey selectivity in planktivorous juvenile fishes associated with floating objects in the western Mediterranean. *Aquaculture Research*, 32(6), 481-490.
49. Devries, D. R., Stein, R. A., & Bremigan, M. T. (1998). Prey selection by larval fishes as influenced by available zooplankton and gape limitation. *Transactions of the American Fisheries Society*, 127(6), 1040-1050.
50. Divita R, Creel M and Sheridan PF (1983) Food of coastal fishes during brown shrimp, *Penaeus azectus*, migration from Texas estuaries (June-July 1981). *U.S. Fishery Bulletin* 81: 396-402.
51. Dörner, H., & Wagner, A. (2003). Size-dependent predator-prey relationships between perch and their fish prey. *Journal of Fish Biology*, 62(5), 1021-1032.
52. Døving, K. B., Sandvig, K., & Kasumyan, A. (2009). Ligand-specific induction of endocytosis in taste receptor cells. *Journal of Experimental Biology*, 212(1), 42-49.
53. Eggers, D. M. (1977). The nature of prey selection by planktivorous fish. *Ecology*, 58(1), 46-59.
54. El Moghraby, A. I., & Abd el Rahman, A. (1984). Food and feeding habits of *Labeo niloticus* (Pisces, Cyprinidae) in Jebel Aulia reservoir, Sudan. *Hydrobiologia*, 110(1), 327-332.
55. Emmanuel, M., Neethu, G. P., Sreekanth, G. B., & Pramod Kiran, R. B. (2019). Food and feeding habits of *Etroplus suratensis* (Bloch, 1790) in Vellayani Lake, Kerala. *Journal of Aquatic Biology and Fisheries*, 7, 120-126.
56. Erzini, K., Gonçalves, J. M., Bentes, L., & Lino, P. G. (1997). Fish mouth dimensions and size selectivity in a Portuguese longline fishery. *Journal of Applied Ichthyology*, 13(1), 41-44.
57. Esmaeili, H. R., & Johal, M. S. (2015). Food and feeding habits of silver carp, *Hypophthalmichthys molitrix* (Val., 1844) in Gobindsagar Reservoir, India. *International Journal of Aquatic Biology*, 3(4), 225-235.
58. Fanelli, E., Principato, E., Monfardini, E., Da Ros, Z., Scarcella, G., Santojanni, A., & Colella, S. (2022). Seasonal Trophic Ecology and Diet Shift in the Common Sole *Solea solea* in the Central Adriatic Sea. *Animals*, 12(23), 3369.

59. Finger, T. E. (1997). Evolution of taste and solitary chemoreceptor cell systems. *Brain, Behavior and Evolution*, 50(4), 234-243.
60. Frid, A., & Marliave, J. (2010). Predatory fishes affect trophic cascades and apparent competition in temperate reefs. *Biology Letters*, 6(4), 533-536.
61. Froese, R., & Pauly, D. (2011). FishBase. World Wide Web Electronic Publications. Available from <http://www.fishbase.org> [accessed 04 June 2011].
62. Froese, R., & Pauly, D. (2023) FishBase. World Wide Web Electronic Publications. Available from <http://www.fishbase.org> [accessed 02 April 2023].
63. Gardner, M. B. (1981). Mechanisms of size selectivity by planktivorous fish: a test of hypotheses. *Ecology*, 62(3), 571-578.
64. Gerhart, D. J., Bondura, M. E., & Commito, J. A. (1991). Inhibition of sunfish feeding by defensive steroids from aquatic beetles: structure activity relationships. *Journal of Chemical Ecology*, 17, 1363-1370.
65. Gerking, S.D. (1994). Feeding ecology of fish. San Diego, California: Academic Press.
66. Giles, N., Street, M., & Wright, R. M. (1990). Diet composition and prey preference of tench, *Tinca tinca* (L.), common bream, *Abramis brama* (L.), perch, *Perca fluviatilis* L. and roach, *Rutilus rutilus* (L.), in two contrasting gravel pit lakes: potential trophic overlap with wildfowl. *Journal of Fish Biology*, 37(6), 945-957.
67. Gill, A. B. (2003). The dynamics of prey choice in fish: the importance of prey size and satiation. *Journal of Fish Biology*, 63, 105-116.
68. Graeb, B. D., Dettmers, J. M., Wahl, D. H., & Cáceres, C. E. (2004). Fish size and prey availability affect growth, survival, prey selection, and foraging behavior of larval yellow perch. *Transactions of the American Fisheries Society*, 133(3), 504-514.
69. Greene, C. H. (1986). Patterns of prey selection: implications of predator foraging tactics. *The American Naturalist*, 128(6), 824-839.
70. Guedes, A. P. P., & Araújo, F. G. (2008). Trophic resource partitioning among five flatfish species (Actinopterygii, Pleuronectiformes) in a tropical bay in south-eastern Brazil. *Journal of Fish Biology*, 72(4), 1035-1054.
71. Gulbrandsen, J. (1991). Functional response of Atlantic halibut larvae related to prey density and distribution. *Aquaculture*, 94(1), 89-98.
72. Hambright, K. D. (1991). Experimental analysis of prey selection by largemouth bass: role of predator mouth width and prey body depth. *Transactions of the American Fisheries Society*, 120(4), 500-508.
73. Hansen, A., & K. Reutter. (2004). Chemosensory systems in fish: structural, functional and ecological aspects. In G., Von der Emde, J. Mogdans, & B. G. Kapoor, (Eds.). *The senses of fish: adaptation for the reception of natural stimuli* (pp. 58-89). Narosa Publishing house, Kluwer Academic Publishers and Springer-Verlag, Dordrecht.
74. Hansen, M. J., & Wahl, D. H. (1981). Selection of small *Daphnia pulex* by yellow perch fry in Oneida Lake, New York. *Transactions of the American Fisheries Society*, 110(1), 64-71.
75. Hasan, M. R., & Macintosh, D. J. (1992). Optimum food particle size in relation to body size of common carp, *Cyprinus carpio* L., fry. *Aquaculture Research*, 23(3), 315-325.
76. Heithaus, M. R., Frid, A., Wirsing, A. J., & Worm, B. (2008). Predicting ecological consequences of marine top predator declines. *Trends in Ecology & Evolution*, 23(4), 202-210.
77. Helfman, G. S., Collette, B. B., Facey, D. E., & Bowen, B. W. (2009). *The Diversity of Fishes: Biology, Evolution, and Ecology*. John Wiley & Sons.
78. Hellawell, J. M., & Abel, R. (1971). A rapid volumetric method for the analysis of the food of fishes. *Journal of Fish Biology*, 3(1), 29-37.
79. Holzman, R., & Genin, A. (2005). Mechanisms of selectivity in a nocturnal fish: a lack of active prey choice. *Oecologia*, 146, 329-336.
80. Horká, P., Sychrová, O., Horký, P., Slavík, O., Švátora, M., & Petrusek, A. (2017). Feeding habits of the alien brook trout *Salvelinus fontinalis* and the native brown



- trout *Salmo trutta* in Czech mountain streams. *Knowledge & Management of Aquatic Ecosystems*, 418, 1-11.
81. Hortle, M. E., & White, R. W. G. (1980). Diet of *Pseudaphritis urvillii* (Cuvier & Valenciennes) (Pisces: Bovichthyidae) from south-eastern Tasmania. *Marine and Freshwater Research*, 31(4), 533-539.
82. Howell, T., Laurenson, L. J., Myers, J. H., & Jones, P. L. (2004). Spatial, temporal and size-class variation in the diet of estuary perch (*Macquaria colonorum*) in the Hopkins River, Victoria, Australia. *Hydrobiologia*, 515(1-3), 29-37.
83. Hunt, S. L., Mulligan, T. J., & Komori, K. (1999). Oceanic feeding habits of Chinook salmon, *Oncorhynchus tshawytscha*, off northern California. *Fishery Bulletin-national Oceanic and Atmospheric Administration*, 97, 717-721.
84. Hurlbert, S. H. (1978). The measurement of niche overlap and some relatives. *Ecology*, 59(1), 67-77.
85. Hyatt, K.D. (1979). Feeding strategy. In W.S., Hoar, D.J., Randall, & J.R Brett (Eds.). *Fish physiology- Bioenergetics and growth* (Vol. 8, pp. 71-119). New York London: Academic Press.
86. Hynes, H. B. N. (1950). The food of freshwater sticklebacks (*Gasterosteus aculeatus* and *Pygosteus pungitius*) with a review of methods used in studies of the food of fishes. *Journal of Animal Ecology* 19, 36-58.
87. Hyslop, E. J. (1980). Stomach contents analysis—a review of methods and their application. *Journal of Fish Biology*, 17(4), 411-429.
88. Ishimaru, Y., Okada, S., Naito, H., Nagai, T., Yasuoka, A., Matsumoto, I., & Abe, K. (2005). Two families of candidate taste receptors in fishes. *Mechanisms of development*, 122(12), 1310-1321.
89. Islam, M. S., & Tanaka, M. (2006). Ontogenetic dietary shift of Japanese sea bass during larva-juvenile transition in Ariake Bay. *Marine Ecology Progress Series*, 323, 305-310.
90. Ivlev, V.S. (1961). *Experimental ecology of the feeding of fishes*. New Haven: Yale University Press.
91. Jackson, J. B., Kirby, M. X., Berger, W. H., Bjorndal, K. A., Botsford, L. W., Bourque, B. J., ... & Warner, R. R. (2001). Historical overfishing and the recent collapse of coastal ecosystems. *Science*, 293(5530), 629-637.
92. Jakubaviciūtė, E., Bergström, U., Ekløf, J. S., Haenel, Q., & Bourlat, S. J. (2017). DNA metabarcoding reveals diverse diet of the three-spined stickleback in a coastal ecosystem. *PLoS One*, 12, 10, e0186929.
93. Jamabo, N. A., & Maduako, N. C. (2015). Food and feeding habits of *Mugil cephalus* (Linnaeus, 1758) in Elechi creek, Niger Delta, Nigeria. *International Journal of Fisheries and Aquaculture*, 7(3), 25-29.
94. Jobling, M. (1995). *Environmental biology of fishes*. London: Chapman & Hall.
95. Jones, K. A. (1984). Temperature dependent attraction by goldfish to a chemical feeding cue presented alone and in combination with heated water. *Physiology & behavior*, 33(4), 509-515.
96. Joseph, P. S., & Mohan Joseph, M. (1988). Feeding habits of the Pearl-Spot *Etroplus suratensis* (Bloch) in the Nethravati-Gurpur estuary, Mangalore. In *Proceedings of the First Indian Fisheries Forum* (pp. 203-206). India: Asian Fisheries Society.
97. Kamal, M.Y. (1967). Studies on the food and alimentary canal of the Indian Major Carps. II. *Labeorohita* (Ham) and III. *Cirrhinus mrigala* (Ham). *Indian Journal of Fisheries*, 14, 24-47.
98. Kamukuru, A. T., & Mgaya, Y. D. (2004). The food and feeding habits of blackspot snapper, *Lutjanus fulviflamma* (Pisces: Lutjanidae) in shallow waters of Mafia Island, Tanzania. *African Journal of Ecology*, 42(1), 49-58.
99. Kasumyan, A. O. (1997). Gustatory reception and feeding behavior in fish. *Journal of Ichthyology*, 37(1), 72-86.
100. Kasumyan, A. O., & DÖving, K. B. (2003). Taste preferences in fishes. *Fish and Fisheries*, 4(4), 289-347.
101. Kawakami, E., & Vazzoler, G. (1980). Metodográfico e estimativa de índice alimentar aplicado no estudo de alimentação de peixes. *Boletim do Instituto Oceanográfico*, 29, 205-207.

102. Keppeler, F. W., Lanés, L. E. K., Rolon, A. S., Stenert, C., Lehmann, P., Reichard, M., & Maltchik, L. (2015). The morphology–diet relationship and its role in the coexistence of two species of annual fishes. *Ecology of Freshwater Fish*, 24(1), 77-90.
103. Khadka, R. B., & Rao, T. R. (1986). Prey size selection by common carp (*Cyprinus carpio* var. *communis*) larvae in relation to age and prey density. *Aquaculture*, 54(1-2), 89-96.
104. Khan, R. A., & Qayyum Siddiqui, A. (1973). Food selection by *Labeorohita* (Ham.) and its feeding relationship with other major carps. *Hydrobiologia*, 43, 429-442.
105. Khan, W., Naqvi, S. M. H. M., Khan, H. U., Rafiq, M., Ahmad, B., Noor, A., ... & Shadman, M. (2021). Feeding habit of Brown trout (*Salmo truttafario*) in upper parts of river Swat, Pakistan. *Brazilian Journal of Biology*, 82. <https://doi.org/10.1590/1519-6984.239219>
106. Knutsen, G. M., & Tilseth, S. (1985). Growth, development, and feeding success of Atlantic cod larvae *Gadus morhua* related to egg size. *Transactions of the American Fisheries Society*, 114(4), 507-511.
107. Kress, W. J., García-Robledo, C., Uriarte, M., & Erickson, D. L. (2015). DNA barcodes for ecology, evolution and conservation. *Trends in Ecology & Evolution*, 30, 25–35.
108. Krishna, P. V., Panchakshari, V., & Prabhavathi, K. (2016). Feeding Habits and Stomach Contents of Asian seabass *Latescalcarifer* from Nizampatnam Coast. *International Journal of Advanced Research*, 4(4), 168-172.
109. Krumme, U., Keuthen, H., Barletta, M., Villwock, W., & Saint-Paul, U. (2005). Contribution to the feeding ecology of the predatory wingfin anchovy *Pterengraulis atherinoides* (L.) in north Brazilian mangrove creeks. *Journal of Applied Ichthyology*, 21(6), 469-477.
110. Labropoulou, M., Machias, A., & Tsimenides, N. (1999). Habitat selection and diet of juvenile red porgy, *Pagrus pagrus* (Linnaeus, 1758). *Fishery Bulletin*, 97(3), 495-507.
111. Lalit, K., Sharma, B. K., Sharma, S. K., Upadhyay, B., & Mishra, V. (2015). Food and feeding habits of *Catla catla* (Hamilton) from lake Udaisagar, Udaipur. *International Journal of Fauna and Biological Studies*, 2(5), 06-08.
112. Lavanya, D., Ramalingaiah, D., Suguna, T., Raveendra Kumar Reddy, D., & Madhavi, K. (2018). Food and feeding ecology of *Mugil cephalus* from Krishnapatnam and Mypadu Coasts of Nellore District, Andhra Pradesh, India. *International Journal of Current Microbiology and Applied Sciences*, 7(4), 2616-2630.
113. Lazzaro, X. (1987). A review of planktivorous fishes: their evolution, feeding behaviours, selectivities, and impacts. *Hydrobiologia*, 146, 97-167.
114. Lehtiniemi, M., Hakala, T., Saesmaa, S., & Viitasalo, M. (2007). Prey selection by the larvae of three species of littoral fishes on natural zooplankton assemblages. *Aquatic Ecology*, 41(1), 85-94.
115. Leonard, J. W., & Leonard, F. A. (1949). An analysis of the feeding habits of rainbow trout and lake trout in Birch Lake, Cass County, Michigan. *Transactions of the American Fisheries Society*, 76(1), 301-314.
116. Lima, S. L. (1998). Stress and decision-making under the risk of predation: recent developments from behavioral, reproductive, and ecological perspectives. *Advances in the Study of Behaviour*, 27(8), 215-290.
117. Lima-Júnior, S.E. (2000). Dieta e condição de *Pimelodus maculatus* (Osteichthyes, Pimelodidae) nos rios Piracicaba e Mogi-Guaçu, SP, Rio Claro, 2000 (*Doctoral dissertation, Dissertação de Mestrado, Instituto de Biociências, UNESP, Campus de Rio Claro*).
118. Lima-Junior, S. E., & Goitein, R. (2001). A new method for the analysis of fish stomach contents. *Acta Scientiarum*, 23(2), 421-424.
119. Limburg, K. E., Pace, M. L., Fischer, D., & Arend, K. K. (1997). Consumption, selectivity, and use of zooplankton by larval striped bass and white perch in a seasonally pulsed estuary. *Transactions of the American Fisheries Society*, 126(4), 607-621.
120. Lin, S. Y. (1935). Life history of WaanUe, *Ctenopharyngodon idellus* (Cuv. & Val.). *Lingnan Science Journal*, 14, 271-274.
121. Liu, Z., & Uiblein, F. (1996). Prey detectability mediates selectivity in a

- zooplanktivorous cyprinid  
(*Alburnus alburnus* (L.)). *Sitzungsber. Abt. 1*, 203, 3-13.
122. Lubzens, E., Sagie, G., Minkoff, G., Meragelman, E., & Schneller, A. (1984). Rotifers (*Brachionus plicatilis*) improve growth of carp (*Cyprinus carpio*) larvae. *Bamidgeh*, 36(2), 41-46.
123. Lückstädt, C., & Reiti, T. (2002). Investigations on the feeding behavior of juvenile milkfish (*Chanoschanos Forsskal*) in brackishwater lagoons on South Tarawa, Kiribati. *Verhandlungen der Gesellschaft für Ichthyologie*, 3, 37-43.
124. Luczkovich, J. J., Norton, S. R., & Gilmore, R. G. (1995). The influence of oral anatomy on prey selection during the ontogeny of two percoid fishes, *Lagodon rhomboides* and *Centropomus undecimalis*. *Environmental Biology of Fishes*, 44, 79-95.
125. Lukoschek, V., & McCormick, M. I. (2001). Ontogeny of diet changes in a tropical benthic carnivorous fish, *Parupeneus barberinus* (Mullidae): relationship between foraging behaviour, habitat use, jaw size, and prey selection. *Marine Biology*, 138, 1099-1113.
126. Lundvall, D., Svanbäck, R., Persson, L., & Byström, P. (1999). Size-dependent predation in piscivores: interactions between predator foraging and prey avoidance abilities. *Canadian Journal of Fisheries and Aquatic Sciences*, 56 (7), 1285-1292.
127. Macdonald, J. S., & Green, R. H. (1983). Redundancy of variables used to describe importance of prey species in fish diets. *Canadian Journal of Fisheries and Aquatic Sciences*, 40(5), 635-637.
128. Mahesh, V., Ambarish, G. P., & Nair, R. J. (2018). Stomach Content Analysis Techniques in Fishes. In *ICAR Sponsored Winter School on Recent Advances in Fishery Biology Techniques for Biodiversity Evaluation and Conservation* (pp. 104-115). Kochi.
129. Malhotra, Y. R., & Langer, S. (1993). Feeding of some fish larvae as determined by prey characteristics. *Journal of the Indian Institute of Science*, 73(5), 457-461.
130. Manko, P. (2016). Stomach content analysis in freshwater fish feeding ecology. *University of Prešov*, ISBN 978-80-555-1613-4.
131. Manly, B. F. J., Miller, P., & Cook, L. M. (1972). Analysis of a selective predation experiment. *The American Naturalist*, 106(952), 719-736.
132. Martin, R. A., Hammerschlag, N., Collier, R. S., & Fallows, C. (2005). Predatory behaviour of white sharks (*Carcharodon carcharias*) at Seal Island, South Africa. *Journal of the Marine Biological Association of the United Kingdom*, 85(5), 1121-1136.
133. Matsumoto, K., & Kohda, M. (2001). Differences in gill raker morphology between two local populations of a benthophagous filter-feeding fish, *Goniistius zonatus* (Cheilodactylidae). *Ichthyological Research*, 48, 269-273.
134. Meng, L. (1993). Estimating food requirements of striped bass larvae: an energetics approach. *Transactions of the American Fisheries Society*, 122(2), 244-251.
135. Metcalf, C., Pezold, F., & Crump, B. G. (1997). Food habits of introduced rainbow trout (*Oncorhynchus mykiss*) in the upper Little Missouri River drainage of Arkansas. *The Southwestern Naturalist*, 42(2), 148-154.
136. Mittelbach, G. G., & Persson, L. (1998). The ontogeny of piscivory and its ecological consequences. *Canadian Journal of Fisheries and Aquatic Sciences*, 55(6), 1454-1465.
137. Mohan, M. V., & Sankaran, T. M. (1988). Two new indices for stomach content analysis of fishes. *Journal of Fish Biology*, 33(2), 289-292.
138. Mohanty, B. P., Mahanty, A., Ganguly, S., Mitra, T., Karunakaran, D., & Anandan, R. (2019). Nutritional composition of food fishes and their importance in providing food and nutritional security. *Food Chemistry*, 293, 561-570.
139. Mondal, A., Chakravorty, D., Mandal, S., Bhattacharyya, S. B., & Mitra, A. (2015). Feeding ecology and prey preference of grey mullet, *Mugil cephalus* (Linnaeus, 1758) in extensive brackish water farming system. *Journal of Marine Science: Research and Development*, 6(1), 1-5.
140. Mondol, M. M. R., Rahman, M. M., Ahamed, F., Sarker, M. A. A., Subba, B. R., & Hossain, M. Y. (2013). Diet and feeding habits of

- Cyprinus carpio* in relation with water quality of integrated rice-fish farming ecosystem. *Our Nature*, 11(2), 138-151.
141. Mookerji, N., & Rao, T. R. (1993). Patterns of prey selection in rohu (*Labeorohita*) and singhi (*Heteropneustes fossilis*) larvae under light and dark conditions. *Aquaculture*, 118(1-2), 85-104.
  142. Mookerji, N., & Rao, T. R. (1995). Prey capture success, feeding frequency and daily food intake rates in rohu, *Labeorohita* (Ham.) and singhi, *Heteropneustes fossilis* (Bloch) larvae. *Journal of Applied Ichthyology*, 11(1-2), 37-49.
  143. Motta, P.J., & Wilga, C.D. (2001). Advances in the study of feeding behaviors, mechanisms, and mechanics of sharks. In T.C., Tricas, & S.H., Gruber, (Eds.). *The behavior and sensory biology of elasmobranch fishes: an anthology in memory of Donald Richard Nelson*. (vol 20., pp. 131-156). Springer, Dordrecht.
  144. Murase, H., Tamura, T., Kiwada, H., Fujise, Y., Watanabe, H., Ohizumi, H., ... & Kawahara, S. (2007). Prey selection of common minke (*Balaenoptera acutorostrata*) and Bryde's (*Balaenoptera edeni*) whales in the western North Pacific in 2000 and 2001. *Fisheries Oceanography*, 16(2), 186-201.
  145. Nakano, S., & Murakami, M. (2001). Reciprocal subsidies: dynamic interdependence between terrestrial and aquatic food webs. *Proceedings of the National Academy of Sciences*, 98(1), 166-170.
  146. Natarajan, A.V., & Jhingran, A.G. (1961). Index of preponderance- a method of grading the food elements in stomach analysis of fishes. *Indian Journal of Fisheries*, 8, 54-59.
  147. Nath, S. R., Beraki, T., Abraha, A., Abraham, K., & Berhane, Y. (2015). Gut content analysis of Indian mackerel (*Rastrelliger kanagurta*). *Journal of Aquaculture and Marine Biology*, 3(1), 1-5.
  148. Nelson, J.S. (2006). *Fishes of the World*. New Jersey: Wiley.
  149. Nikolsky, G.V. (1961). The causes of fluctuations in fish numbers. *Voprosy Ikhtologii*, 1(4).
  150. Nilsson, P. A. (2001). Predator behaviour and prey density: evaluating density-dependent intraspecific interactions on predator functional responses. *Journal of Animal Ecology*, 70(1), 14-19.
  151. Nilsson, P. A., & Brönmark, C. (2000). Prey vulnerability to a gape-size limited predator: behavioural and morphological impacts on northern pike piscivory. *Oikos*, 88(3), 539-546.
  152. Nilsson, P. A., Brönmark, C., & Pettersson, L. B. (1995). Benefits of a predator-induced morphology in crucian carp. *Oecologia*, 104, 291-296.
  153. Northcote, T. G. (1954). Observations on the comparative ecology of two species of fish, *Cottus asper* and *Cottus rhotheus*, in British Columbia. *Copeia*, 1954(1), 25-28.
  154. Nunn, A. D., Harvey, J. P., & Cowx, I. G. (2007). The food and feeding relationships of larval and 0+ year juvenile fishes in lowland rivers and connected waterbodies. I. Ontogenetic shifts and interspecific diet similarity. *Journal of Fish Biology*, 70(3), 726-742.
  155. O'Brien, W. J., Kettle, D., & Riessen, H. (1979). Helmets and invisible armor: structures reducing predation from tactile and visual planktivores. *Ecology*, 60(2), 287-294.
  156. Oike, H., Nagai, T., Furuyama, A., Okada, S., Aihara, Y., Ishimaru, Y., ... & Abe, K. (2007). Characterization of ligands for fish taste receptors. *Journal of Neuroscience*, 27(21), 5584-5592.
  157. Opuszynski, K. (1972). Use of phytophagous fish to control aquatic plants. *Aquaculture*, 1, 61-74.
  158. Ortaz, M. (2001). Diet seasonality and food overlap in fishes of the upper Orituco stream, northern Venezuela. *Revista de Biología Tropical*, 49(1), 191-197.
  159. Oscoz, J., Leunda, P. M., Campos, F., Escala, M. D. C., & Miranda, R. (2005). Diet of 0+ brown trout (*Salmo trutta* L., 1758) from the river Erro (Navarra, north of Spain). *Limnetica*, 24(3-4), 319-325.
  160. Panchakshari, V. (2016). Strategies and diet composition of food and feeding of Asian seabass *Latescalcarifer* from Krishna Estuarine Region, Andhra Pradesh, India. *International Journal of Fisheries and Aquatic Studies*, 4(4), 186-189.



161. Parma, A. M., & Deriso, R. B. (1990). Dynamics of age and size composition in a population subject to size-selective mortality: effects of phenotypic variability in growth. *Canadian Journal of Fisheries and Aquatic Sciences*, 47(2), 274-289.
162. Pauly, D., Christensen, V., Gu  nette, S., Pitcher, T. J., Sumaila, U. R., Walters, C. J., ... & Zeller, D. (2002). Towards sustainability in world fisheries. *Nature*, 418(6898), 689-695.
163. Pepe-Victoriano, R., Aravena-Ambrosetti, H., Huanacuni, J. I., M  ndez-Abarca, F., Godoy, K., &   lvarez, N. (2022). Feeding Habits of *Sarda chiliensis chiliensis* (Cuvier, 1832) in Northern Chile and Southern Per  . *Animals*, 12(7), 930.
164. Pinkas, L. (1971). Food habits of albacore, bluefin tuna, and bonito in California waters. *Fishery Bulletin US*, 152, 1-139.
165. Pinnegar, J. K., Jennings, S., O'brien, C. M., & Polunin, N. V. C. (2002). Long-term changes in the trophic level of the Celtic Sea fish community and fish market price distribution. *Journal of Applied Ecology*, 377-390.
166. Ponton, D., & M  ller, R. (1990). Size of prey ingested by whitefish, *Coregonus* sp., larvae. Are *Coregonus* larvae gape-limited predators? *Journal of Fish Biology*, 36(1), 67-72.
167. Pothoven, S.A., Vanderploeg, H.A., Cavaletto, J.F., Krueger, D.M., Mason, D.M., & Brandt, S.B. (2007). Alewife planktivory controls the abundance of two invasive predatory cladocerans in Lake Michigan. *Freshwater Biology*, 52, 561-573.
168. Prisco, A. R., De La Rosa, S. B. G., & Astrarloo, J. M. D. (2001). Feeding ecology of flatfish juveniles (*Pleuronectiformes*) in Mar Chiquita coastal lagoon (Buenos Aires, Argentina). *Estuaries*, 24(6), 917-925.
169. Priya, P.S.P.V., Ramalingaiah, D., Mohan, R. D., Anupama, R. R., & Ganesh, G. (2020). Study on food and feeding habits of *Etiroplussuratensis* (Bloch, 1790) from Sarvepalli reservoir of Nellore district, Andhra Pradesh. *Journal of Entomology and Zoology Studies*, 8(4), 2349-2351
170. Pusineri, C., Magnin, V., Meynier, L., Spitz, J., Hassani, S., & Ridoux, V. (2007). Food and feeding ecology of the common dolphin (*Delphinus delphis*) in the oceanic Northeast Atlantic and comparison with its diet in neritic areas. *Marine Mammal Science*, 23(1), 30-47.
171. Rasool, N., Jan, U., & Shah, G. M. (2012). Feeding habits and diet composition of Brown trout (*Salmo trutta fario*) in the upper streams of Kashmir Valley. *International Journal of Scientific and Research Publications*, 2(12), 1-8.
172. Rehage, J. S., Barnett, B. K., & Sih, A. (2005). Foraging behaviour and invasiveness: do invasive *Gambusia* exhibit higher feeding rates and broader diets than their noninvasive relatives? *Ecology of Freshwater Fish*, 14(4), 352-360.
173. Rezende, C. F., Caramaschi, E. P., & Mazzoni, R. (2008). Fluxo de energia em comunidades aqu  ticas, com   nfase em sistemas l  ticos. *Oecologia Brasiliensis*, 12(4), 626-639.
174. Rockstrom, J., Steffen, W., Noone, K., Persson, A., Chapin III, F. S., Lambin, E. F., ... & Foley, J. A. (2009). A safe operating space for humanity: identifying and quantifying planetary boundaries that must not be transgressed could help prevent human activities from causing unacceptable environmental change, argue Johan Rockstrom and colleagues. *Nature*, 461(7263), 472-476.
175. Rom  n-Valencia, C. (1998). Alimentaci  n y reproducci  n de *Creagrutus brevipinnis* (Pisces: Characidae) en alto Cauca, Colombia. *Revista de Biolog  a Tropical*, 46(3), 783-789.
176. Ross, S. T. (1986). Resource partitioning in fish assemblages: a review of field studies. *Copeia*, 352-388.
177. Sahtout, F., Boualleg, C., Kaouachi, N., Kh  lifi, N., Menasria, A., & Bensouilah, M. (2018). Feeding habits of *Cyprinus carpio* in Fom El-Khanga Dam, Souk-Ahras, Algeria. *Aquaculture, Aquarium, Conservation & Legislation*, 11(2), 554-564.
178. Saikia, S. K., & Das, D. N. (2009). Feeding ecology of common carp (*Cyprinus carpio* L.) in a rice-fish culture system of the Apatani plateau (Arunachal Pradesh, India). *Aquatic Ecology*, 43, 559-568.

179. Saliu, J. K. (2002). Size, sex and seasonal dynamics in the dietary composition of *Brycinus nurse* (Pisces: Characidae), from Asa reservoir, Ilorin, Nigeria. *Revista de Biología Tropical*, 233-238.
180. Šantić, M., Jardas, I., & Pallaoro, A. (2005). Feeding habits of horse mackerel, *Trachurus trachurus* (Linnaeus, 1758), from the central Adriatic Sea. *Journal of Applied Ichthyology*, 21(2), 125-130.
181. Santos, J., & Borges, T. (2001). Trophic relationships in deep-water fish communities off Algarve, Portugal. *Fisheries Research*, 51(2-3), 337-341.
182. Schael, D. M., Rudstam, L. G., & Post, J. R. (1991). Gape limitation and prey selection in larval yellow perch (*Perca flavescens*), freshwater drum (*Aplodinotus grunniens*), and black crappie (*Pomoxis nigromaculatus*). *Canadian Journal of Fisheries and Aquatic Sciences*, 48(10), 1919-1925.
183. Scharf, F. S., Buckel, J. A., Juanes, F., & Conover, D. O. (1998). Predation by juvenile piscivorous bluefish (*Pomatomus saltatrix*): the influence of prey to predator size ratio and prey type on predator capture success and prey profitability. *Canadian Journal of Fisheries and Aquatic Sciences*, 55(7), 1695-1703.
184. Scharnweber, K., Watanabe, K., Syväranta, J., Wanke, T., Monaghan, M. T., & Mehner, T. (2013). Effects of predation pressure and resource use on morphological divergence in omnivorous prey fish. *BMC Evolutionary Biology*, 13(132), 1-12.
185. Scheffer, M., Carpenter, S., & de Young, B. (2005). Cascading effects of overfishing marine systems. *Trends in Ecology & Evolution*, 20(11), 579-581.
186. Schoener, T. W. (1968). The Anolis lizards of Bimini: resource partitioning in a complex fauna. *Ecology*, 49(4), 704-726.
187. Schoener, T. W. (1970). Nonsynchronous spatial overlap of lizards in patchy habitats. *Ecology*, 51(3), 408-418.
188. Schulte, B. A., & Bakus, G. J. (1992). Predation deterrence in marine sponges: laboratory versus field studies. *Bulletin of Marine Science*, 50(1), 205-211.
189. Sih, A., & Christensen, B. (2001). Optimal diet theory: when does it work, and when and why does it fail? *Animal Behaviour*, 61(2), 379-390.
190. Simpfendorfer, C. A., Heupel, M. R., White, W. T., & Dulvy, N. K. (2011). The importance of research and public opinion to conservation management of sharks and rays: a synthesis. *Marine and Freshwater Research*, 62(6), 518-527.
191. Sirois, P., & Dodson, J. J. (2000). Critical periods and growth-dependent survival of larvae of an estuarine fish, the rainbow smelt *Osmerus mordax*. *Marine Ecology Progress Series*, 203, 233-245.
192. Spataru, P., & Gophen, M. (1985). Feeding behaviour of silver carp *Hypophthalmichthys molitrix* Val. and its impact on the food web in Lake Kinneret, Israel. *Hydrobiologia*, 120, 53-61.
193. Stephens, D. W., & Krebs, J. R. (1986). *Foraging theory* (Vol. 6). Princeton University Press.
194. Sujatha, K., Joice, A. A., & Kumaar, P. S. (2013). Total protein and lipid content in edible tissues of fishes from Kasimodu fish landing centre, Chennai, Tamilnadu. *European Journal of Experimental Biology*, 3(5), 252-257.
195. Svenning, M. A., Klemetsen, A., & Olsen, T. (2007). Habitat and food choice of Arctic charr in Linnévatn on Spitsbergen, Svalbard: The first year-round investigation in a High Arctic lake. *Ecology of Freshwater Fish*, 16(1), 70-77.
196. Takeda, M., Takii, K. (1992). Gustation and nutrition in fishes: application to aquaculture. In T.J., Hara, (Eds.). *Fish Chemoreception. Fish & Fisheries Series*, (Vol 6, pp. 271-287). Dordrecht: Springer. [https://doi.org/10.1007/978-94-011-2332-7\\_13](https://doi.org/10.1007/978-94-011-2332-7_13)
197. Tararam, A. S., Wakabara, Y., & Equi, M. B. (1993). Hábitos alimentares de onze espécies da megafauna benthica da plataforma continental de Ubatuba, SP. *Publicacao Especial do Instituto Oceanografico. Universidade de Sao Paulo*, (10), 159-167.
198. Taylor, M. D., Fielder, D. S., & Suthers, I. M. (2006). Spatial and ontogenetic variation in the diet of wild and stocked mullet. *Journal of Fish Biology*, 68(2), 389-400.

- (*Argyrosomus japonicus*, Sciaenidae) in Australian estuaries. *Estuaries and Coasts*, 29(5), 785-793.
199. Tucker, J.W. (1998). Marine Fish Culture. Boston, Massachusetts: Kluwer Academic Publishers.
200. Turesson, H., Persson, A., & Brönmark, C. (2002). Prey size selection in piscivorous pikeperch (*Stizostedion lucioperca*) includes active prey choice. *Ecology of Freshwater Fish*, 11(4), 223-233.
201. Wahl, C. M., Mills, E. L., McFarland, W. N., & DeGisi, J. S. (1993). Ontogenetic changes in prey selection and visual acuity of the yellow perch, *Perca flavescens*. *Canadian Journal of Fisheries and Aquatic Sciences*, 50(4), 743-749.
202. Wahl, D. H., & Stein, R. A. (1988). Selective predation by three esocids: the role of prey behavior and morphology. *Transactions of the American Fisheries Society*, 117(2), 142-151.
203. Wainwright, P. C. (1988). Morphology and ecology: functional basis of feeding constraints in Caribbean labrid fishes. *Ecology*, 69(3), 635-645.
204. Wainwright, P. C. (1996). Ecological explanation through functional morphology: the feeding biology of sunfishes. *Ecology*, 77(5), 1336-1343.
205. Wainwright, P. C., & Richard, B. A. (1995). Predicting patterns of prey use from morphology of fishes. *Environmental Biology of Fishes*, 44, 97-113.
206. Wang, S., Tang, J. P., Su, L. H., Fan, J. J., Chang, H. Y., Wang, T. T., ... & Yang, Y. (2019). Fish feeding groups, food selectivity, and diet shifts associated with environmental factors and prey availability along a large subtropical river, China. *Aquatic Sciences*, 81, 1-18.
207. Wanzenböck, J. (1995). Changing handling times during feeding and consequences for prey size selection of 0+ zooplanktivorous fish. *Oecologia*, 104(3), 372-378.
208. Watkins, C.E., Shireman, J.V., Watkins, C. E., Shireman, J. V., Rottmann, R. W., & Colle, D. E. (1981). Food habits of fingerling grass carp. *The Progressive Fish-Culturist*, 43(2), 95-97.
209. Webb, P. W. (1978). Partitioning of energy into metabolism and growth. *Ecology of Freshwater Fish Production*, 184-214.
210. Webb, P. W. (1986). Effect of body form and response threshold on the vulnerability of four species of teleost prey attacked by largemouth bass (*Micropterus salmoides*). *Canadian Journal of Fisheries and Aquatic Sciences*, 43(4), 763-771.
211. Welker, M. T., Pierce, C. L., & Wahl, D. H. (1994). Growth and survival of larval fishes: roles of competition and zooplankton abundance. *Transactions of the American Fisheries Society*, 123(5), 703-717.
212. Werner, E. E. (1974). The fish size, prey size, handling time relation in several sunfishes and some implications. *Journal of the Fisheries Board of Canada*, 31(9), 1531-1536.
213. Whitear, M. (1971). Cell specialization and sensory function in fish epidermis. *Journal of Zoology*, 163(2), 237-264.
214. Wilkens, M.A., Wettring, B., Wagner, E., Wojtenek, W. & Russell, D. (2001). Prey detection in selective plankton feeding by the paddle fish: is the electric sense sufficient? *Journal of Experimental Biology*, 204(8), 1381-1389
215. Williams, M. J. (1981). Methods for analysis of natural diet in portunid crabs (Crustacea: Decapoda: Portunidae). *Journal of Experimental Marine Biology and Ecology*, 52(1), 103-113.
216. Windell, J.T. & Bowen, S.H. (1978). Methods of study of fish based on analysis of stomach contents. In T., Benegal (Ed.). *Methods Assessment of Fish Production in Freshwaters* (pp. 219-226). Oxford: Blackwell Scientific.
217. Winemiller, K.O., Agostinho, A.A., & Caramaschi, E.P. (2008). Fish ecology in tropical streams. In D., Dudgeon (Ed.). *Tropical Stream Ecology* (pp 107-146). San Diego: Elsevier Academic Press.
218. Wootton, R.J. (1998). *Ecology of teleost fishes*. Dordrecht: Kluwer Academic Publishers.
219. Yashpal, M., Kumari, U., Mittal, S. & Mittal, A.K. (2009). Morphological specialization of the buccal cavity in relation to the food and feeding habit of a carp *Cirrhinus mrigala*: a scanning electron microscopic investigation. *Journal of Morphology*, 270 (6), 714- 728
220. Yeon, I. J., Hong, S. H., Cha, H. K., & Kim, S. T. (1999). Feeding habits of *Raja pulchra* in the Yellow Sea. *Bulletin of National Fisheries Research Development Institute Korea*, 57, 1-11.

221. Yúfera, M., & Darias, M. J. (2007a). The onset of exogenous feeding in marine fish larvae. *Aquaculture*, 268(1-4), 53-63.
222. Yúfera, M., & Darias, M. J. (2007b). Changes in the gastrointestinal pH from larvae to adult in Senegal sole (*Solea senegalensis*). *Aquaculture*, 267(1-4), 94-99.
223. Zaki, S., Jayabalan, N., Al-Marzouqi, A., Al-Kiyumi, F., & Al-Anboori, I. (2021). Reproduction and feeding of the Indian Oil Sardine *Sardinella longiceps* Val. from Mahout along the Arabian Sea Coast of Oman. In L.A., Jawad (Ed.). *The Arabian Seas: Biodiversity, Environmental Challenges and Conservation Measures* (pp. 499-518). Springer, Cham.
224. Zaret, T.M. (1980). *Predation and freshwater communities*. New Haven: Yale University Press.
-